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#### (54) INSECTICIDAL PROTEINS

(75) Inventors: **Yan Gao**, Research Triangle Park, NC

(US); **Jared Conville**, Research Triangle Park, NC (US); **Jeng Shong Chen**, Research Triangle Park, NC (US)

(73) Assignee: Syngenta Participations AG, Basel

(CH)

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- (52) U.S. CI. CPC ............ *C12N 15/8286* (2013.01); *C07K 14/325* (2013.01)

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#### (58) Field of Classification Search

None

See application file for complete search history.

#### (56) References Cited

#### U.S. PATENT DOCUMENTS

#### FOREIGN PATENT DOCUMENTS

WO WO2011/041256 A2 4/2011

#### OTHER PUBLICATIONS

Syngenta Participations AG, WO2011/041256 International Search Report, issued Jun. 9, 2011.

\* cited by examiner

Primary Examiner — David H Kruse (74) Attorney, Agent, or Firm — Gregory W. Warren

#### (57) ABSTRACT

Improved compositions and methods for controlling pests are disclosed. In particular, novel engineered Cry1Ba (eCry1Ba) proteins having improved toxicity to lepidopteran insect pests are provided. By substituting at least one amino acid in domain I of a Cry1Ba protein an engineered Cry1Ba protein having substantially altered insecticidal properties is designed. Further, a method of making the engineered Cry1Ba proteins and methods of using the ecry1Ba nucleic acid sequences, for example in transgenic plants to express eCry1B proteins to confer protection from insect damage are disclosed.

19 Claims, 6 Drawing Sheets

% Identity	99 99 100 99	neneiinavsnhsaqmdllpdariedslciaegnnidpfvsastvqtginiagrilgvlgvpfagqlasfysfl	vgelwprgrdqweiflehveqlinqqitenarntalarlqglgdsfrayqqsledwlenrddartrsvlytqyialeldf         .h         .h         lnamplfairnqevpllmvyaqaanlhllllrdaslfgsefgltsqeiqryyerqvertrdysdycvewyntglnslrgt	
Matches	1227 1227 1227 1228 1228	aegnnidpf	arlqglgds:	
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Start		einavsnhs	dqweiflehv	
SEQ ID No.	4 4 4 4 0 7 6 7 8 9 0	1 mtsnrknen 1 1 1		61 61 61 61 61
Cry Name	CrylBal CrylBa2 CrylBa4 CrylBa5 CrylBa5		Н Н	
Pos	Ref 1 1 2 2 4 2 2 5 4 6 5 6 6 5 4 6 6 6 6 6 6 6 6 6 6 6 6	Crylbal Crylba2 Crylba3 Crylba4 Crylba5 Crylba5	CrylBal CrylBa2 CrylBa4 CrylBa4 CrylBa5 CrylBa6 CrylBa6	CrylBa3 CrylBa4 CrylBa5 CrylBa6

Figure 1A

. U Q	aswvrynqfirdlilgvldlvalfpsydtrtypintsaqltrevytdaigatgynmasmnwynnnapsfsaleaaair  g g g  hlldflegltifsassrwsntrhmtywrghtigsrpiggglntsthgatntsinpvtlrfasrdvyrtesyagvllwg  hlldflegltifsassrwsntrhmtywrghtigsrpiggglntsthgatntsinpvtlrfasrdvyrtesyagvllwg  r  r  hlldflegltifsassrwsntrhmtywrghtigsrpiggglntsthgatntsinpvtlrfasrdvyrtesyagvllwg  r  r  r  faffvergdttunfiflitmnsgdelkygnfvrrafttpfffqiqdiirtsiqglsgngevyidkieiipvtatfe	a · · · · · · · · · · · · · · · · · · ·	CrylBal CrylBa2 CrylBa3 CrylBa4 CrylBa4 CrylBa1 CrylBa3 CrylBa3 CrylBa3 CrylBa3 CrylBa3 CrylBa3 CrylBa3 CrylBa3 CrylBa3 CrylBa1 CrylBa3
		199	ry1Ba2 -v1Ba3
561	dffvsrggttvnnfrflrtmnsgdelkygnfvrrafttpftftqiqdiirtsiqglsgngevyidkieiipvtatfe	61 vd	ry1Ba1
561		481	ry1Ba6
481 561 vd 561		481	ry1Ba5
481 481 561 vd 561		481	ry1Ba4
481 481 561 vd 561 vd		481	ry1Ba3
481 481 481 561		481	ry1Ba2
481 481 481  481  561 	thrsadrtntigpnritqipmvkaselpqgttvvrgpgftggdilrrtntggfgpirvtvngpltgryrigfryast	1 ys	ry1Ba1
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401 481.ys 481 481 481 561		401	ry1Ba5
401 481 YS 481 481 481 481 561		401	ry1Ba4
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401 401 401 481. Ys 481. Ys 481. Ys 481. Ys 761. Y	epihgvptvrfnftnpqnisdrgtanysqpyespglqlkdsetelppetterpnyesyshrlshigiilqsrvnvpv	iΣ	ry1Ba1
401 i.y 401 i.y 401 i.y 401 i.y 481 i.y 481 i.y 61		$\sim$	ry1Ba6
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Figure 1B

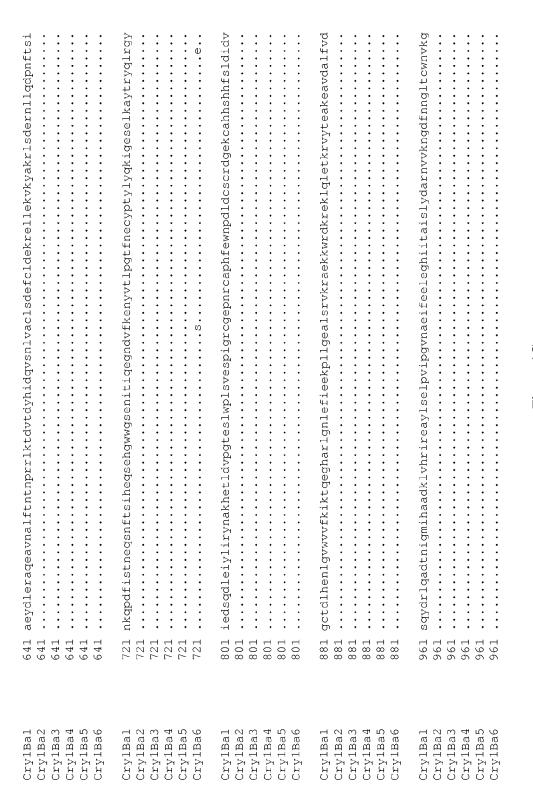


Figure 1C

CrylBal	1041 hvdvqqshhrsdlvıpeweaevsqavrvcpgcgyılrvtaykegygegcvtıheıenntdelkiknreeeevyptdtgtc 1011
Crylba2	1041
Cry1Ba3	1041
$\mathtt{Cry1Ba4}$	1041
Cry1Ba5	1041
Cry1Ba6	1041
Crv1Ba1	1121 ndytahggtagcadaensrnagyedavevdttasvnykptveeetytdyrrdnheevdrgyvnyppypagyvtkeleyfp
Cry1Ba2	1121
Cry1Ba3	1121
CrylBa4	1121
Cry1Ba5	1121
CrylBa6	1121
CrylBal	1201 etdtvwieigetegkfivdsvelllmee
Cry1Ba2	1201
Cry1Ba3	1201
Cry1Ba4	1201
Cry1Ba5	1201
CrylBa6	1201mm

%Matches		54	mtsnrkneneiinavsnhsaqmdllpdariedslciaegnnidpfvsastvqtginiagrilgvlgvpfagqlasfys -md.np.ic.py.clpevev.ggetgytpidislsltqfl.sefgvl	flvgelwpr-grdqweiflehveqlinqqitenarntalarlqglgdsfrayqqsledwlenrddarfrsvlytqyiale gdii.gif.psdavqir.e.fq.isesnlyqi.ae.fre.eadptnpal.eemri.fndmn	α-helix 5  Idflnamplfalrnqevp llmvyaqaanlhllllrdaslfgsefgltsqeiqryyerqvertrdysdycvewyntglnsl saltt.ivq.yq s.vsvv.v.qrw.fdaat.nsr.ndltrlign.t.ha.rerv	$\begin{tabular}{ll} \hline \textbf{Domain II} \\ rgtnaaswvrynqfrrdltlgvldlvalfpsydtrtypintsaqltrevytdaigatgvnmasmnwynnnapsfsaieaa\\ w.pdsrd.ieti.sn sr.vsipvle.fdgsfrg.aggg\\ \hline \end{tabular}$	airsphlldfleqltifsassrwsntrhmtywrghtiqsrpiggglntsthgatntsinpvtlrfasrdvyrtesy sm.i.nsiytdah.geysq.mas.v.fsgpef.fplyg.mg.aapqqrivaqlgqgl.s	agvllwgiylepih-gvptvrfnftnpqnisdrgtany-sqpyespglqlkdsetelppetterpnyesyshrlshigii tl.rr.fni.innqqlsvldgtefaygtss.lp.av.rkstvld.iqnnnv.prqgfvsmf	lqsrvnvpvyswthrsadrtntigpnritqipmvkaselpqgttvvrgpgftggdilrrtntggfgpirvtvn rsgfsnssv.iira.mfiefn.i.pssqlt.stn.gssksp.qistlnit
Matches		989	.dpfvsastvq tgytpid	ılgdsfrayqq .snlyqi.ae	gltsqeiqry 7.fdaat.nsr	<b>II</b>  ltrevytdaio  i	glntsthgat: sgpef.fply	spglqlkdse kstvl	elpqgttvvr n.gssk
Length	1228 aa	1155 aa	Islciaegnni 	rntalarlqg q.ise.	rdaslfgsefv.vqrw	Domain II.	htiqsrpigg .q.mas.v.f	rtany-sqpye .ss.lp.av.r	itqipmvkas [lt.st
End	1228	1155	lpdaried .gge	nggitens r.e.f.	<b>α-helix 5</b> anlhllll sv.	in I alfpsydt Sn.s	rhmtywrg ys.	oqnisdro gtefaygt	tntigpnr n.i.pssc
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SEQ ID NO:	7	51	1 mtsnrknenei 1 -md.np.ic	79 flvgelwpr-g 59 gdii.gif.	ldflna saltt			agvllw t	lq rsgfsnss
Sequence	CrylBal	$\mathtt{Cry1Ab3}$		7 2	158 139	238	318 290	3 8 8 4 8 5 5 4 4 5 5 5 5 5 5 5 5 5 5 5 5	472
Pos	Ref 1	7	CrylBal CrylAb3	CrylBal CrylAb3	CrylBal CrylAb3	CrylBal CrylAb3	CrylBal CrylAb3	CrylBal CrylAb3	CrylBal CrylAb3

Figure 2/

CrylBal 545 gpltqryrigfryastvdfdffvsrggttvnnfrflrtmnsgdelkygnfvrrafttpftftqiqdiirtsigglsgnge CrylAb3 517 asvritnlq.ht.id.rpi.qgn.sassn.qs.s.rtvgn.sngssvftl.ahvfnsgn.	CrylBal 625 vyidkieiipvtatfeaeydleraqeavnalftntnprrlktdvtdyhidqvsnlvaclsdefcldekrellekvkyakr CrylAb3 597r.fv.aevkess.qigeeek	CrylBal 705 lsdernllqdpnftsinkqpdfistneqsnftsiheqsehgwwgsenitiqegndvfkenyvtlpgtfnecyptylygki CrylAb3 677rgrgr.l	CrylBal 785 geselkaytryqlrgyiedsqdleiylirynakhetldvpgteslwplsvespigrcgepnrcaphfewnpdldcscrdg CrylAb3 739 dk	CrylBal 865 ekcahhshhfsldidvgctdlhenlgvwvvfkiktqegharlgnlefieekpllgealsrvkraekkwrdkreklqletk CrylAb3 794	CrylBal 945 rvyteakeavdalfvdsqydrlqadtnigmihaadklvhrireaylselpvipgvnaeifeeleghiitaislydarnvv CrylAb3 873 iksnar.sr.si	CrylBal       1025 kngdfnngltcwnvkghvdvq-qshhrsdlvipeweaevsqavrvcpgcgyilrvtaykegygegcvtiheienntdelk         CrylAb3       953see.nnv.v.v.v	CrylBal 1104 fknreeeevyptdtgtcndytahqgtagcadacnsrnagyedayevdttasvnykptyeeetytdvrrdnhceydr CrylAb3 1033 .s.cvnn.vtqeeyeytr.dgsnssvpad.asakagpsn.	CrylBal 1180 gyvnyppvpagyvtkeleyfpetdtvwieigetegkfivdsvelllmee CrylAb3 1107 .gd.t.lkktt	Figure 2B
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#### INSECTICIDAL PROTEINS

#### **CROSS-REFERENCE**

This is a national stage application under 35 U.S.C. §371 of 5 International Application No. PCT/US10/50369, filed on Sep. 27, 2010, which is entitled to the benefit of U.S. Provisional Application No. 61/247,986, filed on Oct. 2, 2009, both of which are incorporated herein by reference in their entirety.

#### STATEMENT REGARDING ELECTRONIC SUBMISSION OF A SEQUENCE LISTING

A Sequence Listing in ASCII text format, submitted under 37 C.F.R. §1.821, entitled "72633\_US\_REG\_ORG\_P\_1\_ 22Apr2013\_SubstituteSeqList\_ST25", 169 KB in size, generated on Apr. 22, 2013 and filed via EFS-Web is provided in lieu of a paper copy. This Sequence Listing is hereby incorporated by reference into the specification for its disclosure.

#### FIELD OF THE INVENTION

The invention relates to the fields of protein engineering, plant molecular biology and pest control. More particularly, the invention relates to novel engineered Cry1Ba proteins and 25 nucleic acid sequences whose expression results in the engineered Cry1Ba proteins, and methods of making and methods of using the engineered Cry1Ba proteins and corresponding nucleic acid sequences to control insects.

#### BACKGROUND

Bacillus thuringiensis (Bt) Cry proteins (also called δ-endotoxins or Cry toxins) are proteins that form a crystalline matrix in Bacillus that are known to possess insecticidal 35 activity when ingested by certain insects. Over 180 holotype Cry proteins in 58 families have been identified and named. The various Cry proteins have been classified based upon their spectrum of activity and sequence homology. Prior to activity (Hofte and Whitely, 1989, Microbiol. Rev. 53:242-255), but more recently a new nomenclature was developed which systematically classifies the Cry proteins based on amino acid sequence homology rather than insect target specificities (Crickmore et al. 1998, Microbiol. Molec. Biol. 45 Rev. 62:807-813).

Most Cry proteins active against lepidopteran insects are formed in the crystalline matrix as 130-140 kDa protoxins. In lepidopteran insects, the alkaline pH of the gut solubilizes the crystal and then gut proteases process the protoxin to toxic 50 proteins of approximately 60-70 kDa. Processing of the protoxin to toxin has been reported to proceed by removal of both N- and C-terminal amino acids with the exact location of processing being dependent on the specific Cry protein and the specific insect gut fluids involved (Ogiwara et al., 1992, J. 55 Invert. Pathol. 60:121-126). The proteolytic activation of a Cry protoxin can play a significant role in determining its specificity.

The three dimensional structure for several Cry proteins has been elucidated. The Cry3A protein, which is active 60 against coleopteran insects, has three structural domains: the N-terminal domain I, from residues 58-290, consists of 7 alpha-helices, domain II, from residues 291-500, contains three beta-sheets in a so-called Greek key-conformation, and the C-terminal domain III, from residues 501-644, is a beta- 65 sandwich in a so-called jellyroll conformation. The three dimensional structure for the lepidopteran active Cry1Aa

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toxin has also been solved (Grochulski et al., 1995, J. Mol. Biol. 254:447-464). The Cry1Aa toxin also has three domains: the N-terminal domain I, from residues 33-253, domain II from residues 265-461, and domain III from residues 463-609 with an additional outer strand in one of the  $\beta$ -sheets formed by residues 254-264. If the Cry3A and Cry1Aa structures are projected on other Cry1 sequences, domain I runs from about amino acid residue 28 to 260, domain II from about 260 to 460 and domain III from about 460 to 600. See, Nakamura et al., Agric. Biol. Chem. 54(3): 715-724 (1990); Li et al., Nature 353: 815-821 (1991); Ge et al., J. Biol. Chem. 266(27): 17954-17958 (1991); and Honee et al., Mol. Microbiol. 5(11): 2799-2806 (1991); each of which are incorporated herein by reference. Thus, it is now known that based on amino acid sequence homology, all Bt Cry proteins have a similar three-dimensional structure comprising three domains.

Based on the structure, a hypothesis has been formulated regarding the structure/function relationship of the Cry pro-20 teins. It is generally thought that domain I, the most N-terminal domain, is primarily responsible for pore formation in the insect gut membrane (Gazit & Shai, 1993, Appl. Environ. Microbiol. 57:2816-2820), domain II is primarily responsible for interaction with a gut receptor thereby determining toxin specificity (Ge et al., 1991, J. Biol. Chem. 32:3429-3436) and domain III, the most C-terminal domain, is most likely involved with protein stability (Li et al. 1991, supra) as well as having a regulatory impact on ion channel activity (Chen et al., 1993, PNAS 90:9041-9045). Domain III has also been 30 implicated in determining specificity (U.S. Pat. No. 6,204, 246, herein incorporated by reference). Swapping domain III between lepidopteran-active toxins, such as by in vivo recombination between the coding regions, can result in changes in specific activity. Binding experiments using such hybrids have shown that domain III is involved in binding to putative receptors of target insects, suggesting that domain III may have some impact on specificity through a role in receptor

The toxin portions of Bt Cry proteins are also characterized 1990, the major classes were defined by their spectrum of 40 by having five conserved blocks across their amino acid sequence (Hofte & Whiteley, supra). Conserved block 1 (CB1) comprises approximately 29 amino acids. Conserved block 2 (CB2) comprises approximately 67 amino acids. Conserved block 3 (CB3) comprises approximately 48 amino acids. Conserved block 4 (CB4) comprises approximately 10 amino acids. Conserved block 5 (CB5) comprises approximately 12 amino acids. The sequences before and after these five conserved blocks are highly variable and thus are designated the "variable regions," V1-V6. Domain I of a Bt Cry protein typically comprises a C-terminal portion of variable region 1, a complete conserved block 1, an entire variable region 2, and the N-terminal 52 amino acids of conserved block 2. Domain II typically comprises approximately the C-terminal 15 amino acids of conserved block 2, a variable region 3, and approximately the N-terminal 10 amino acids of conserved block 3. Domain III typically comprises approximately the C-terminal 38 amino acids of conserved block 3, variable region 4, conserved block 4, variable region 5, and conserved block 5. The Cry1 lepidopteran active toxins, among other Cry proteins, have a variable region 6 with approximately 1-3 amino acids lying within domain III.

Several Cry proteins, for example Cry1Ab, Cry1Ac, Cry1F and Cry2Ba have been expressed in transgenic crop plants and exploited commercially to control certain lepidopteran insect pests. For example, transgenic corn hybrids expressing a Cry1Ab protein have been available commercially for over 10 years. The Cry1Ab protein in these corn hybrids targets

primarily European corn borer (Ostrinia nubilalis), the major lepidopteran pest in the US Corn Belt.

One concern raised regarding the deployment of transgenic crops expressing a Cry protein is whether insect pests will become resistant to the Cry protein. Insects have proven 5 capable of developing resistance to Cry protein-containing products. Resistance in diamondback moth (Plutella xylostella) and other vegetable pests to commercial Bt microbial sprays, used extensively in organic farming, has developed in several parts of the world. One recent incidence of field resistance in a fall armyworm (Spodoptera frugiperda) population exposed to transgenic corn expressing Cry1F protein has been documented on the island of Puerto Rico (Storer et al. 2010. J. Econ. Entomol. 103:1031-1038). However, there have been no cases of any field failures in the United States associated 15 with resistant field populations of corn or cotton pests exposed to transgenic crops since 1996 when transgenic crops expressing Cry proteins were first introduced.

The seed industry, university researchers and the US Environmental Protection Agency have worked together to 20 develop management plans to help mitigate the onset of insect resistance. They are based primarily on a high dose and refuge strategy. A high dose strategy for European corn borer in corn, for example, is to use corn hybrids that express high enough levels of a Cry protein to kill even partially resistant European 25 corn borers. The underlying hypothesis is that killing partially resistant ECB and preventing their mating greatly delays the development of resistance. The success of a high dose strategy depends in part on the specific activity of the Cry toxin to European corn borer and how much of that Cry toxin can be 30 expressed in the transgenic corn plant. For example, the higher the specific activity of a Cry toxin to a pest, the less amount of Cry toxin is required to be expressed in a transgenic plant to achieve a high dose strategy. Because Cry1Ab is very toxic to European corn borer larvae (i.e. high specific 35 activity) levels of expression of Cry1 Ab that are achievable in transgenic plants easily places such corn hybrids in a high dose category.

Other possible ways to mitigate resistance development include pyramiding multiple Cry proteins in the same trans- 40 genic crop plant or replacing existing mature products with new products that produce different Cry proteins. For example, as the current Cry1Ab corn hybrid market matures, new products may be introduced that have Cry proteins other than Cry1Ab or other Cry proteins in addition to Cry1Ab. It 45 would be beneficial for proteins that replace Cry1Ab to have the same or similar specific activity to European corn borer as Cry1Ab.

One candidate Cry toxin to replace Cry1Ab may be a Cry1Ba toxin. The holotype Cry1Ba toxin was first described 50 by Brizzard et al. in 1988 (Nuc. Acids Res. 16:2723-2724). Subsequently, five other Cry1Ba toxins have been identified with each having about 99% identity to the holotype toxin. Cry1Ba toxins have been reported to have activity against certain lepidopteran pests, such as cabbage butterfly (Pieris 55 brassicae), diamondback moth (Plutella xylostella), Egyptian cotton leafworm (Spodoptera littoralis), beet armyworm (Spodoptera exigua) and European corn borer (Ostrinia nubilalis). However, Cry1Ba has been reported to be greater than 2-fold less active against European corn borer than Cry1Ab 60 (See for example, U.S. Pat. No. 5,628,995) and has been reported to have no activity against other major corn pests, for example corn earworm (Helicoverpa zea) (See for example, Karim et al. 2000. Pestic. Biochem. Physiol. 67: 198-216) and NAFTA populations of fall armyworm (Spodoptera fru- 65 giperda) (See for example, Monnerat et al. 2006. Appl. Environ. Microbiol. 72:7029-7035). One reason that Cry1Ba is

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not as active as Cry1Ab against at least European corn borer may be due to its lower solubility properties. Thus, there is a need to improve the specific activity of Cry1Ba against at least European corn borer and possibly expand its spectrum of activity to increase its potential as a replacement for Cry1Ab in transgenic corn.

The spectrum of insecticidal activity of an individual Cry toxin from Bt may be quite narrow, with a given Cry toxin being active against only a few species within an Order. For instance, the Cry3A protein is known to be very toxic to the Colorado potato beetle, Leptinotarsa decemlineata, but has very little or no toxicity to related beetles in the genus Diabrotica (Johnson et al., 1993, J. Econ. Entomol. 86:330-333). In addition, small variations in amino acid sequence within a Cry protein class can impact insecticidal activity. For example, von Tersch et al. (1991, Appl. Environ. Microbiol. 57:349-358) demonstrated that Cry1Ac proteins varying by seven amino acids showed significant differences in their spectrum of insecticidal activity. Although considered primarily lepidopteran-active toxins. Cry 1Ba toxins have also been reported to be active against certain coleopteran insects pests including Colorado potato beetle (Leptinotarsa decemlineata), cottonwood leaf beetle (Chrysomela scripta) and coffee berry borer (Hypothenemus hampei).

Specificity of the Cry proteins is the result of the efficiency of the various steps involved in producing an active toxin protein and its subsequent interaction with the epithelial cells in the insect mid-gut. To be insecticidal, most known Cry proteins must first be ingested by the insect and proteolytically activated to form an active toxin. Activation of the insecticidal crystal proteins is a multi-step process. After ingestion, the crystals must first be solubilized in the insect gut. Once solubilized, the Cry proteins are activated by specific proteolytic cleavages. The proteases in the insect gut can play a role in specificity by determining where the Cry protein is processed. Once the Cry protein has been solubilized and processed it binds to specific receptors on the surface of the insects' mid-gut epithelium and subsequently integrates into the lipid bilayer of the brush border membrane. Ion channels then form disrupting the normal function of the midgut eventually leading to the death of the insect. There are stark differences in the solubility properties of the toxin portions of Cry proteins.

Certain lepidopteran-active Cry proteins have been engineered in attempts to improve specific activity or to broaden the spectrum of insecticidal activity. For example, the silk moth (*Bombyx mori*) specificity domain from Cry1Aa was moved to Cry1Ac, thus imparting a new insecticidal activity to the resulting chimeric protein (Ge et al. 1989, PNAS 86: 4037-4041). Also, Bosch et al. 1998 (U.S. Pat. No. 5,736, 131), created a new lepidopteran-active toxin by substituting domain III of Cry1E with domain III of Cry1C thus producing a Cry1E-Cry1C hybrid toxin with a broader spectrum of lepidopteran activity.

There remains a need to design new and effective pest control agents that provide an economic benefit to farmers and that are environmentally acceptable. Needed are proteins with substantially altered properties, such as the engineered Cry1Ba proteins of the invention, that have greater specific activity than native Cry1Ba proteins against at least European corn borer, a major pest of corn in the United States, that may become resistant to existing insect control agents. Furthermore, engineered Cry1Ba proteins whose use minimizes the burden on the environment, as through transgenic plants, are desirable.

By increasing the specific activity of Cry1Ba to at least European corn borer, less Cry1Ba protein should be needed to

be expressed in a maize plant therefore reducing the possible negative impacts of Cry1Ba on the plant. In addition, the increased specific-activity allows for use of the engineered Cry1Ba in a high dose strategy for ECB.

#### **SUMMARY**

In view of these needs, it is an object of the invention to provide novel engineered Cry1Ba (eCry1Ba) proteins having substantially altered properties that are improved over and 10 distinct from native Cry1Ba proteins, particularly biochemical properties associated with the insecticidal activity to lepidopteran pests of corn, including but not limited to such pests as European corn borer (ECB; Ostrinia nubilalis), corn earworm (CEW; Helicoverpa zea), southwestern corn borer 15 (SWCB; Diatraea grandiosella), sugarcane borer (SCB; Diatraea saccharalis), soybean looper (SBL; Pseudoplusia includens), velvet bean caterpillar (VBC; Anticarsia gemmatalis), and the like. By substituting amino acids at key identified positions in a native Cry1Ba protein sequence or 20 wild-type Cry1Ba protein as defined herein, in accordance with the present invention, an eCry1Ba protein having substantially altered solubility and/or insecticidal properties compared to native Cry1Ba is designed. The invention is further drawn to nucleic acid sequences encoding the 25 eCry1Ba proteins, and to compositions and formulations containing the eCry1Ba proteins, which are capable of inhibiting the ability of insect pests to survive, grow and reproduce, or of limiting insect-related damage or loss to crop plants. The invention is further drawn to a method of making the eCry1Ba 30 proteins and to methods of using the eCry1Ba proteins, for example in transgenic plants to confer protection from insect damage. The substantially altered properties of the eCry1Ba proteins of the invention allow for their use in a high dose strategy against at least ECB while requiring expression lev- 35 els in corn plants that are readily achievable.

The novel eCry1Ba proteins described herein are highly active against insects. For example, the eCry1Ba proteins of the invention can be used to improve control of economically important insect pests such as ECB or CEW without negatively impacting activity against other important corn pests such as SWCB and SCB. The eCry1Ba proteins of the invention can be used singly or in combination with other insect control strategies to confer maximal pest control efficiency with minimal environmental impact. Transgenic plants 45 expressing the eCry1Ba proteins of the invention provide a means by which growers can control major lepidopteran pests of crops, for example without limitation corn and sugar cane.

According to one aspect, the present invention includes an engineered Cry1Ba (eCry1Ba) protein comprising a mutation 50 at one or more amino acid positions in domain I, whereby the engineered Cry1Ba protein has improved solubility and/or insecticidal activity against at least European corn borer (*Ostrinia nubilalis*) when compared to a native or wild-type Cry1Ba protein.

In another aspect, the mutation at one or more amino acid positions is located in alpha-helix 4 or alpha-helix 5 of domain I.

In a further aspect, the mutation is at an amino acid position corresponding to position 150, 178, 189 or 199 of SEQ ID 60 NO: 2

In yet another aspect, the mutation is at position 150, 178, 189 or 199 of SEQ ID NO: 5.

In another aspect, the mutation is at a position corresponding to amino acids 2 and 150; or amino acids 2, 150 and 178; 65 or amino acids 2, 150 and 189; or amino acids 2, 150 and 199, of SEQ ID NO: 5.

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In still another aspect, the mutation is at amino acids 2 and 150; or amino acids 2, 150 and 178; or amino acids 2, 150 and 189; or amino acids 2, 150 and 199, of SEQ ID NO: 5.

In one aspect, the invention includes an engineered Cry1Ba (eCry1Ba) protein comprising the amino acid sequence of SEQ ID NO: 6, wherein X at position 2 is any amino acid and a) X at position 150 is Pro, Phe, Trp or Lys, and X at position 189 is Leu and at position 199 is Ser; or b) X at position 189 is Ser when X at position 150 is Lys; or c) X at position 199 is Lys when X at position 150 is Lys.

In another aspect, the eCry1Ba protein of the invention comprises the amino acid sequence of SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9 or SEQ ID NO: 10.

In yet another aspect, an eCry1Ba protein of the invention has activity against lepidopteran or coleopteran insects, particularly lepidopteran insects. Examples of such lepidopteran insects include but are not limited to European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean caterpillar.

The invention also includes other variant Cry1Ba proteins (vCry1Ba) wherein a tyrosine (Tyr) or histidine (His) at position 150 (Y150 or H150) is substituted with an amino acid other than Tyr or His. In one aspect, the amino acid that is substituted for Y150 or H150 is Lys, Phe, Trp, Pro, Thr, Leu, Ala, Val, Ser, Arg, Gly or Asp.

In another aspect, the invention includes a vCry1Ba protein that comprises SEQ ID NO: 3.

In still another aspect, the invention includes a vCry1Ba protein wherein a Tyr or His at position 150 is substituted with an amino acid other than Tyr or His and also has a valine (Val) at position 81 (V81) substituted with an amino acid other than Val; or an alanine (Ala) at position 155 (A155) and a methionine (Met) at position 178 (M178) substituted with amino acids other than Ala or Met, respectively. In another aspect, the Val at position 81 (V81) is substituted with a tryptophan (Trp) (V81W). In yet another aspect, a variant Cry1Ba protein of the invention comprises SEQ ID NO: 11.

In one aspect, the invention includes a vCry1Ba protein with the Y150 substituted with any other amino acid and, wherein an Ala at position 155 (A155) is substituted with an aspartic acid (Asp) (A155D) and a Met at position 178 (Met178) is substituted with a serine (Ser) (M178S). In another aspect, a variant Cry1Ba protein of the invention comprises SEQ ID NO: 12.

The vCry1Ba proteins of the invention have insecticidal activity against lepidopteran or coleopteran insects, particularly lepidopteran insects. Such lepidopteran insects include without limitation European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean caterpillar. However, such vCry1Ba proteins may not have increased activity compared to the wild-type Cry1Ba protein against such pests.

In another aspect, the present invention includes a nucleic acid that encodes an engineered Cry1Ba (eCry1Ba) protein of the invention or a variant Cry1Ba protein (vCry1Ba) of the invention.

The present invention also includes a chimeric gene comprising a heterologous promoter sequence operatively linked to a nucleic acid that encodes an eCry1Ba protein or a vCry1Ba protein. The present invention also includes a recombinant vector comprising such a chimeric gene. Further, the present invention includes a transgenic non-human host cell comprising such a chimeric gene. A transgenic host cell according to this aspect of the invention includes without limitation a bacterial cell or a plant cell. Such a transgenic plant cell may be a maize cell or a sugar cane cell.

The present invention further provides a transgenic plant comprising such a plant cell. The eCry1Ba proteins or vCry1Ba proteins are useful for expressing in any transgenic plant where susceptible insect pests are a problem. In another aspect of the invention, progeny plants comprising a nucleic 5 acid of the invention from any generation of a transgenic plant and a propagule comprising a nucleic acid of the invention from any generation of a transgenic plant are included. In another aspect, the transgenic plant is a maize plant or a sugar cane plant. In yet another aspect, the propagule is a seed, a 10 sette or a cutting.

The invention also includes an insecticidal composition comprising an effective insect-controlling amount of an eCry1Ba protein or a vCry1Ba protein according to the invention and additionally an acceptable agricultural carrier. Such 15 agricultural carriers may be, for example, a sparyable formulation or a transgenic plant.

In another aspect, the present invention provides a method of producing a eCry1Ba protein or a vCry1Ba protein that is active against insects, comprising: (a) obtaining a host cell 20 comprising a chimeric gene, which itself comprises a heterologous promoter sequence operatively linked to a nucleic acid of the invention; and (b) expressing the nucleic acid in the transgenic host cell, which results in at least one protein that is active against insects.

In a further aspect, the present invention provides a method of producing an insect-resistant transgenic plant, comprising introducing a nucleic acid of the invention into a plant thereby producing a transgenic plant, wherein the nucleic acid causes the expression of a eCry1Ba protein or a vCry1Ba protein in 30 the transgenic plant in an effective amount to control insects. In yet another aspect, the insects are lepidopteran or coleopteran insects. Such lepidopteran insects include without limitation European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet 35 bean caterpillar.

In another aspect, the invention includes a method of making an engineered Cry1Ba (eCry1Ba) protein comprising a) identifying a Cry1Ba protein having a domain I; b) substituting at least one native amino acid at a location in domain I 40 with at least one other amino acid; and c) obtaining the eCry1Ba protein so produced, wherein the eCry1Ba has improved solubility and/or insecticidal activity against at least European corn borer when compared to a native Cry 1Ba protein or a wild-type Cry1Ba. In still another aspect, the 45 native amino acid is located in alpha-helix 4 or alpha-helix 5 of domain I.

In still another aspect, the invention includes a method of controlling a lepidopteran insect comprising contacting the insect with an effective amount of an eCry1Ba protein or a 50 tein. vCry1Ba protein of the invention. According to another aspect, such lepidopteran insects include without limitation European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean cater-

Preferably, the eCry1Ba protein or vCry1Ba protein is delivered to the insects orally. In one aspect, the proteins are delivered orally through a transgenic plant comprising a nucleic acid sequence that expresses a eCry1Ba or vCry1Ba protein of the invention.

The present invention further provides a method of controlling insects wherein a transgenic plant comprising a nucleic acid encoding an eCry1Ba protein or vCry1Ba protein further comprises a second nucleic acid sequence or multiple nucleic acid sequences that encode at least one other 65 pesticidal principle. In one aspect, the second nucleic acid sequence encodes a Cry protein different then the eCry1Ba or

vCry1Ba proteins of the invention, those that encode a Vegetative Insecticidal Protein, such as those disclosed in U.S. Pat. Nos. 5,849,870 and 5,877,012, which are incorporated herein by reference, or those that encode a pathway for the production of a non-proteinaceous pesticidal principle. In another aspect of the invention, the second insecticidal principle is a Vip3 protein.

Yet another aspect of the invention is the provision of a method of providing a grower with an improved means of controlling European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean caterpillar comprising supplying or selling to the grower transgenic propagules comprising a nucleic acid that encodes an eCry1Ba protein having a mutation at one or more amino acid positions in domain I, the eCry1Ba protein having improved solubility or insecticidal activity against at least European corn borer when compared to a native Cry1Ba protein. In another aspect, the transgenic propagules are seeds, settes or cuttings.

Other aspects and advantages of the present invention will become apparent to those skilled in the art from a study of the following description of the invention and non-limiting examples.

#### BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 shows an alignment of the amino acid sequences of the known native Cry1Ba proteins (SEQ ID NOs:45-50). Amino acids at position 150 are in bold.

FIG. 2 shows an alignment of the amino acid sequences of the native full-length Cry1Ab (SEQ ID NO:51) and Cry1Ba (SEQ ID NO:2). Domains I and II and alpha-helices 4 & 5 of Domain I are shown by arrows.

#### BRIEF DESCRIPTION OF THE SEQUENCES IN THE SEQUENCE LISTING

SEQ ID NO: 1 is a native full-length cry1Ba coding sequence.

SEQ ID NO: 2 is the amino acid sequence of a native full-length Cry1Ba protein.

SEQ ID NO: 3 is a mutated full-length Cry1Ba.

SEQ ID NO: 4 is the cry1Ba-T25 coding sequence.

SEQ ID NO: 5 is the Cry1Ba-T25 wild-type protein.

SEQ ID NO: 6 is the eCry1Ba-X150 protein.

SEQ ID NO: 7 is the eCry1Ba-T2AY150K protein.

SEQ ID NO: 8 is the eCry1Ba-T2AY150KM178S protein.

SEQ ID NO: 9 is the eCry1Ba-T2AY150KL189S protein.

SEQ ID NO: 10 is the eCry1Ba-T2AY150KS199K pro-

SEQ ID NO: 11 is variant Cry1Ba-TM21.

SEQ ID NO: 12 is variant Cry1Ba-TM90.

SEQ ID NO: 13 is a maize-optimized nucleic acid sequence encoding eCry1Ba-T2AY150KL189S protein.

SEQ ID NOs:14-41 are primers useful in the invention.

SEQ ID NO: 42 is a truncated native Cry1Ba.

SEQ ID NO: 43 is variant Cry1Ba-TM69.

SEQ ID NO: 44 is variant Cry1Ba-TM61.

SEQ ID NOs: 45-51 are Cry toxins shown in FIGS. 1 and 60 **2**.

#### **DEFINITIONS**

For clarity, certain terms used in the specification are defined and presented as follows:

"Activity" of an eCry1Ba protein of the invention is meant that the eCry1Ba proteins function as orally active insect

control agents, have a toxic effect, or are able to disrupt or deter insect feeding, which may or may not cause death of the insect. When a eCry1Ba protein of the invention is delivered to the insect, the result is typically death of the insect, or the insect does not feed upon the source that makes the eCry1Ba 5 protein available to the insect.

"Associated with/operatively linked" refer to two nucleic acid sequences that are related physically or functionally. For example, a promoter or regulatory DNA sequence is said to be "associated with" a DNA sequence that codes for an RNA or 10 a protein if the two sequences are operatively linked, or situated such that the regulatory DNA sequence will affect the expression level of the coding or structural DNA sequence.

A "chimeric gene" or "chimeric construct" is a recombinant nucleic acid sequence in which a promoter or regulatory 15 nucleic acid sequence is operatively linked to, or associated with, a nucleic acid sequence that codes for an mRNA or which is expressed as a protein, such that the regulatory nucleic acid sequence is able to regulate transcription or expression of the associated nucleic acid coding sequence. 20 The regulatory nucleic acid sequence of the chimeric gene is not normally operatively linked to the associated nucleic acid sequence as found in nature.

A "coding sequence" is a nucleic acid sequence that is transcribed into RNA such as mRNA, rRNA, tRNA, snRNA, 25 sense RNA or antisense RNA. Preferably the RNA is then translated in an organism to produce a protein.

To "control" insects means to inhibit, through a toxic effect, the ability of insect pests to survive, grow, feed, and/or reproduce, or to limit insect-related damage or loss in crop 30 plants. To "control" insects may or may not mean killing the insects, although it preferably means killing the insects.

As used herein, the term "corn" means *Zea mays* or maize and includes all plant varieties that can be bred with corn, including wild maize species.

"Corresponding to" in the context of the present invention means that when the amino acid sequences of Cry1B proteins are aligned with each other, the amino acids that "correspond to" certain enumerated positions in the present invention are those that align with these positions in the native Cry1Ba 40 toxin (SEQ ID NO: 2), but that are not necessarily in these exact numerical positions relative to the particular Cry1Ba amino acid sequence of the invention. For example, the methionine at position 1 of a truncated Cry1Ba protein (SEQ ID NO: 42) will align with the methionine at position 22 of the 45 full-length Cry1Ba (SEQ ID NO: 2). Therefore, according to the present invention, amino acid 129 of SEQ ID NO: 42 "corresponds to" amino acid number 150 of SEQ ID NO: 2.

To "deliver" a toxin means that the toxin comes in contact with an insect, resulting in toxic effect and control of the 50 insect. The toxin can be delivered in many recognized ways, e.g., orally by ingestion by the insect or by contact with the insect via transgenic plant expression, formulated protein composition(s), sprayable protein composition(s), a bait matrix, or any other art-recognized toxin delivery system.

"Effective insect-controlling amount" means that concentration of toxin that inhibits, through a toxic effect, the ability of insects to survive, grow, feed and/or reproduce, or to limit insect-related damage or loss in crop plants. "Effective insect-controlling amount" may or may not mean killing the insects, 60 although it preferably means killing the insects.

An "engineered Cry1Ba" (eCry1Ba) protein of the invention, refers to a Cry1Ba-derived protein having at least one mutation in domain I which is not known to naturally occur in a Cry1Ba protein. An eCry1Ba protein is not naturally occurring and, by the hand of man, comprises an amino acid sequence that is not identical to a protein known to occur in

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Bacillus thuringiensis. Cry1Ba proteins that have been engineered according to the invention have substantially altered and improved properties compared to native Cry1Ba proteins. Particularly, eCry1Ba proteins of the invention have improved solubility and/or insecticidal activity against at least European corn borer compared to a native Cry1Ba, wild-type Cry1Ba or variant Cry1Ba proteins of the invention.

An "engineered cry1Ba gene" (ecry1Ba) according to this invention, refers to a nucleic acid comprising the coding sequence of an eCry1B protein. The engineered cry1Ba gene can be derived from a native cry1Ba gene or from a synthetic cry1Ba gene.

"Expression cassette" as used herein means a nucleic acid sequence capable of directing expression of a particular nucleic acid sequence in an appropriate host cell, comprising a promoter operably linked to the nucleic acid sequence of interest which is operably linked to termination signals. It also typically comprises sequences required for proper translation of the nucleic acid sequence. The expression cassette comprising the nucleic acid sequence of interest may be chimeric, meaning that at least one of its components is heterologous with respect to at least one of its other components. The expression cassette may also be one that is naturally occurring but has been obtained in a recombinant form useful for heterologous expression. Typically, however, the expression cassette is heterologous with respect to the host, i.e., the particular nucleic acid sequence of the expression cassette does not occur naturally in the host cell and must have been introduced into the host cell or an ancestor of the host cell by a transformation event. The expression of the nucleic acid sequence in the expression cassette may be under the control of a constitutive promoter or of an inducible promoter that initiates transcription only when the host cell is exposed to some particular external stimulus. In the case of a multicellular organism, such as a plant, the promoter can also be specific to a particular tissue, or organ, or stage of develop-

A "gene" is a defined region that is located within a genome and that, besides the aforementioned coding nucleic acid sequence, comprises other, primarily regulatory, nucleic acid sequences responsible for the control of the expression, that is to say the transcription and translation, of the coding portion. A gene may also comprise other 5' and 3' untranslated sequences and termination sequences. Further elements that may be present are, for example, introns.

"Gene of interest" refers to any gene which, when trans50 ferred to a plant, confers upon the plant a desired characteristic such as antibiotic resistance, virus resistance, insect
resistance, disease resistance, or resistance to other pests,
herbicide tolerance, improved nutritional value, improved
performance in an industrial process or altered reproductive
55 capability. The "gene of interest" may also be one that is
transferred to plants for the production of commercially valuable enzymes or metabolites in the plant.

As used herein, the term "grower" means a person or entity that is engaged in agriculture, raising living organisms, such as crop plants, for food or raw materials.

A "gut protease" is a protease naturally found in the digestive tract of an insect. This protease is usually involved in the digestion of ingested proteins.

A "heterologous" nucleic acid sequence is a nucleic acid sequence not naturally associated with a host cell into which it is introduced, including non-naturally occurring multiple copies of a naturally occurring nucleic acid sequence.

A "homologous" nucleic acid sequence is a nucleic acid sequence naturally associated with a host cell into which it is introduced

"Homologous recombination" is the reciprocal exchange of nucleic acid fragments between homologous nucleic acid 5 molecules

"Insecticidal" is defined as a toxic biological activity capable of controlling insects, preferably by killing them.

An "isolated" nucleic acid molecule or protein is a nucleic acid molecule or protein that, by the hand of man, exists apart from its native environment and is therefore not a product of nature. An isolated nucleic acid molecule or protein may exist in a purified form or may exist in a non-native environment such as, for example, a recombinant host cell. For example, a native Cry protein naturally occurring in *Bacillus thuringiensis* is not isolated, but that same Cry protein in a transgenic *Bacillus thuringiensis* strain or a transgenic plant is isolated.

A "native" Cry1Ba protein as used herein, refers to an approximately 140 kDa Bacillus thuringiensis (Bt) 20 coleopteran- or lepidopteran-active protein, for example SEQ ID NO: 2, as well as any truncated lower molecular weight protein derivable from a native Cry1Ba protein that has an amino acid sequence found in nature. The lower molecular weight protein can be obtained by protease cleavage of natu- 25 rally occurring protease recognition sites of the native Cry1Ba protein or by a second translational initiation codon in the same frame as the translational initiation codon coding for the native Cry1Ba protein, for example M22 of SEQ ID NO: 2. The amino acid sequence of a native Cry1Ba protein 30 and the lower molecular weight proteins derived thereof may be found in a protein naturally occurring in Bt. For example, six native Cry1Ba proteins have been named and have the following Genbank accession numbers,

Cry1Ba1=CAA29898 (SEQ ID Cry1Ba2=CAA65003 (SEQ ID NO:46); Cry1Ba3=AAK63251 (SEQ ID NO:47); Cry1Ba4=AAK51084 (SEQ ID NO:48); Cry1Ba5=AB020894 (SEQ IDNO:49); Cry1Ba6=ABL60921 (SEQ ID NO:50). A sequence align- 40 ment of six native Cry1Ba proteins is shown in FIG. 1. A native Cry1Ba protein can be encoded by a native Bt nucleotide sequence as in SEQ ID NO: 1 or by a synthetic codon

A "nucleic acid molecule" or "nucleic acid sequence" is a 45 linear segment of single- or double-stranded DNA or RNA that can be isolated from any source. In the context of the present invention, the nucleic acid molecule or nucleic acid sequence is preferably a segment of DNA.

optimized nucleotide sequence.

A "plant" is any plant at any stage of development, particu- 50 larly a seed plant.

A "plant cell" is a structural and physiological unit of a plant, comprising a protoplast and a cell wall. The plant cell may be in the form of an isolated single cell or a cultured cell, or as a part of a higher organized unit such as, for example, 55 plant tissue, a plant organ, or a whole plant.

"Plant cell culture" means cultures of plant units such as, for example, protoplasts, cell culture cells, cells in plant tissues, pollen, pollen tubes, ovules, embryo sacs, zygotes and embryos at various stages of development.

"Plant material" refers to leaves, stems, roots, flowers or flower parts, fruits, pollen, egg cells, zygotes, seeds, settes, cuttings, cell or tissue cultures, or any other part or product of a plant.

A "plant organ" is a distinct and visibly structured and 65 differentiated part of a plant such as a root, stem, leaf, flower bud, or embryo.

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"Plant tissue" as used herein means a group of plant cells organized into a structural and functional unit. Any tissue of a plant in planta or in culture is included. This term includes, but is not limited to, whole plants, plant organs, plant seeds, tissue culture and any groups of plant cells organized into structural and/or functional units. The use of this term in conjunction with, or in the absence of, any specific type of plant tissue as listed above or otherwise embraced by this definition is not intended to be exclusive of any other type of plant tissue.

A "promoter" is an untranslated DNA sequence upstream of the coding region that contains the binding site for RNA polymerase and initiates transcription of the DNA. The promoter region may also include other elements that act as regulators of gene expression.

A "propagule" is any plant material used for the purpose of plant propagation. For example, without limitation, seeds, and cuttings or settes are propagules of corn and sugar cane, respectively.

A "protoplast" is an isolated plant cell without a cell wall or with only parts of the cell wall.

"Regulatory elements" refer to sequences involved in controlling the expression of a nucleic acid sequence. Regulatory elements comprise a promoter operably linked to the nucleic acid sequence of interest and termination signals. They also typically encompass sequences required for proper translation of the nucleic acid sequence.

As used herein, "specific activity" refers to the amount of protein required to have an insecticidal effect. Therefore, when a first protein has a higher specific activity than a second protein means that it takes a lesser amount of the first protein compared the second protein to have an insecticidal effect on the same percentage of insects.

"Solubility" as used herein refers to the quantity of a native NO:45); 35 Cry1Ba toxin, wild-type Cry1Ba or eCry1Ba toxin or VCry1Ba toxin that can dissolve in a particular liquid, for example a buffer, water or insect gut fluid, under the same environmental conditions. Thus, as used herein, an eCry1Ba toxin has "improved solubility" or an "increase in solubility" compared to a native or wild-type Cry1Ba toxin means that a given volume of liquid can hold a great quantity of an eCry1Ba toxin than a native or wild-type Cry1Ba toxin under the same conditions. According to this invention, native Cry1Ba and wild-type Cry1Ba toxins have low solubility and certain eCry1Ba toxins have high solubility, relative to each other.

"Transformation" is a process for introducing heterologous nucleic acid into a host cell or organism. In particular, "transformation" means the stable integration of a DNA molecule into the genome of an organism of interest.

"Transformed/transgenic/recombinant" refer to a host organism such as a bacterium or a plant into which a heterologous nucleic acid molecule has been introduced. The nucleic acid molecule can be stably integrated into the genome of the host or the nucleic acid molecule can also be present as an extrachromosomal molecule. Such an extrachromosomal molecule can be auto-replicating. Transformed cells, tissues, or plants are understood to encompass not only the end product of a transformation process, but also transgenic progeny thereof. A "non-transformed", "non-transgenic", or "non-recombinant" host refers to a wild-type organism, e.g., a bacterium or plant, which does not contain the heterologous nucleic acid molecule.

A "variant Cry1Ba (vCry1Ba)" protein is a non-native mutant protein that has lower specific activity to at least European corn borer compared to a wild-type Cry1Ba protein of the invention.

A "wild-type Cry1Ba" protein is a non-native mutated protein that has similar insecticidal properties, such as specific activity, against such insects as southwestern corn borer, sugarcane borer or European corn borer, or biochemical properties, such as solubility, as a native Cry1Ba.

Nucleic acids are indicated by their bases by the following standard abbreviations: adenine (A), cytosine (C), thymine (T), and guanine (G) Amino acids are likewise indicated by the following standard abbreviations: alanine (Ala; A), arginine (Arg; R), asparagine (Asn; N), aspartic acid (Asp; D), cysteine (Cys; C), glutamine (Gln; Q), glutamic acid (Glu; E), glycine (Gly; G), histidine (His; H), isoleucine (Ile; 1), leucine (Leu; L), lysine (Lys; K), methionine (Met; M), phenylalanine (Phe; F), proline (Pro; P), serine (Ser; S), threonine (Thr; T), tryptophan (Trp; W), tyrosine (Tyr; Y), and valine (Val; V).

#### DETAILED DESCRIPTION

This invention relates to novel engineered Cry1Ba 20 (eCry1Ba) proteins having substantially altered properties improved over and distinct from native Cry1Ba proteins, particularly biochemical properties associated with the insecticidal activity to lepidopteran pests of corn, including but not limited European corn borer (ECB; Ostrinia nubilalis), corn 25 earworm (CEW; Helicoverpa zea), southwestern corn borer (SWCB; Diatraea grandiosella), sugarcane borer (SCB; Diatraea saccharalis), soybean looper (SBL; Pseudoplusia includens), velvet bean caterpillar (VBC; Anticarsia gemmatalis), and the like. By mutating amino acids at key identified positions in a native Cry1Ba protein sequence, in accordance with the present invention, an eCry1Ba protein having substantially altered solubility and/or insecticidal properties compared to a native Cry1Ba or a wild-type Cry1Ba as defined herein is designed. Nucleic acid sequences that 35 encode the eCry1Ba proteins can be used, for example, in transgenic crop plants to cause the expression of the eCry1Ba proteins to control insect pests such as European corn borer (ECB; Ostrinia nubilalis), corn earworm (CEW; Helicoverpa zea), southwestern corn borer (SWCB; Diatraea grandi- 40 osella), sugarcane borer (SCB; Diatraea saccharalis), soybean looper (SBL; Pseudoplusia includens), velvet bean caterpillar (VBC; Anticarsia gemmatalis), and the like.

In one embodiment, the present invention encompasses an engineered Cry1Ba (eCry1Ba) protein comprising a mutation 45 at one or more amino acid positions in domain I, whereby the eCry1Ba protein has improved solubility and/or insecticidal activity against at least European corn borer when compared to a native or wild-type Cry1Ba protein.

In another embodiment, the mutation at one or more amino 50 acid positions is located in alpha-helix 4 or alpha-helix 5 of domain I. Studies of the structure-function relationship of certain Cry proteins, such as Cry1Aa, Cry1Ab, and Cry1Ac, have included mutagenesis of alpha-helices 4 and 5 of Domain I (Saraswathy et al. 2004, Electron. J. Biotech. 7: 55 178-188). Results of these experiments implicate alpha-helix 4 and 5 in ion channel formation and conductance. It is not clear whether any one mutation or a combination of mutations in Domain I of a Cry protein, particularly Cry1Ba would have an impact on solubility and specific activity. Therefore, 60 Domain I of a Cry1Ba protein, particularly at locations in alpha-helix 4 or 5, was targeted for mutational analysis to determine if the solubility could be improved and/or the specific activity of a native Cry1Ba protein could be increased against a target insect, including European corn borer (ECB), 65 southwestern corn borer (SWCB), sugarcane borer (SCB), corn earworm (CEW), soybean looper (SBL) and velvet bean

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caterpillar (VBC), as well as others. Based on sequence alignment, alpha-helix 4 of Cry1Ba comprises amino acids 143-163 of SEQ ID NO: 2. Alpha-helix 5 makes up a majority of Conserved Block 1 and comprises amino acids 176-199 of SEQ ID NO: 2. The six known native Cry1Ba proteins vary by only one amino acid in alpha-helix 4 at position 150. Four of the six have a tyrosine (Tyr; Y) at position 150 and the other two have a histidine (His; H) at position 150. The instant disclosure now demonstrates that the amino acid at position 150 plays a critical role in the toxicity of a Cry1Ba protein and that mutations in alpha-helix 4 and alpha-helix 5 can have a significant impact on protein solubility, specific activity against a particular pest and increase in spectrum of activity of Cry1Ba.

In another embodiment, the invention encompasses mutations at an amino acid position corresponding to position 150, 178, 189 or 199 of SEQ ID NO: 2. In yet another embodiment, the mutation is at position 150, 178, 189 or 199 of SEQ ID NO: 5.

In still another embodiment, the invention encompasses mutations in Cry1Ba at a position corresponding to amino acids 2 and 150; or amino acids 2, 150 and 178; or amino acids 2, 150 and 189; or amino acids 2, 150 and 199, of SEQ ID NO: 5. In another embodiment, the mutation is at amino acids 2 and 150; or amino acids 2, 150 and 178; or amino acids 2, 150 and 189; or amino acids 2, 150 and 199, of SEQ ID NO: 5.

In one embodiment, the invention encompasses an engineered Cry1Ba (eCry1Ba) protein comprising the amino acid sequence of SEQ ID NO: 6, wherein Xaa at position 2 is any amino acid and a) Xaa at position 150 is Pro, Phe, Trp or Lys, and Xaa at position 189 is Leu and at position 199 is Ser; or b) Xaa at position 189 is Ser when Xaa at position 150 is Lys; or c) Xaa at position 199 is Lys when Xaa at position 150 is Lys.

In another embodiment, the invention encompasses an eCry1Ba protein that comprises the amino acid sequence of SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9 or SEQ ID NO: 10

In yet another embodiment, an eCry1Ba protein of the invention has activity against lepidopteran or coleopteran insects, particularly against lepidopteran insects. Examples of such lepidopteran insects include but are not limited to European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean caterpillar. Engineered Cry1Ba proteins of the invention also have activity against corn earworm, an insect pest for which native Cry1Ba is not active.

In still another embodiment, the invention encompasses an eCry1Ba protein that has at least a 3× higher specific activity than a native Cry1Ba protein against at least European corn borer.

In another embodiment, the invention also encompasses variant Cry1Ba (vCry1Ba) proteins wherein a tyrosine (Tyr) or histidine (His) at position 150 (Y150 or H150) is substituted with an amino acid other than Tyr or His. In one aspect, the amino acid that is substituted for Y150 or H150 is Lys, Phe, Trp, Pro, Thr, Leu, Ala, Val, Ser, Arg, Gly or Asp.

In another embodiment, the invention encompasses a mutated Cry1Ba protein that comprises SEQ ID NO: 3.

In still another embodiment, the invention encompasses a vCry1Ba protein wherein a Tyr or His at position 150 (Y150 or H150) is substituted with an amino acid other than Tyr or His and also has a valine (Val) at position 81 (V81) substituted with an amino acid other than Val; or an alanine (Ala) at position 155 (A155) and a methionine (Met) at position 178 (M178) substituted with amino acids other than Ala or Met, respectively. In another embodiment, the Val at position 81 (V81) is substituted with a tryptophan (Trp) (V81W). In yet

another embodiment, the invention encompasses a variant Cry1Ba protein that comprises SEQ ID NO: 11.

In one embodiment, the invention encompasses a vCry1Ba protein with the Tyr at 150 (Y150) substituted with any other amino acid and, wherein an Ala at position 155 (A155) is substituted with an aspartic acid (Asp) (A155D) and a Met at position 178 (M178) is substituted with a serine (Ser) (M178S). In another embodiment, the invention encompasses a vCry1Ba protein comprising SEQ ID NO: 12.

The vCry1Ba proteins encompassed by the invention have 10 insecticidal activity against a lepidopteran or coleopteran insect. Such a lepidopteran insect includes without limitation European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean caterpillar. The activity of vCry1Ba proteins is typically less than 15 a wild-type Cry1Ba protein of the invention. One advantage of such variant Cry1Ba proteins is their usefulness in situations where high specific activity is not required. The skilled person will recognize other uses and advantages of such variant Cry1Ba proteins.

The insect controlling properties of the eCry1Ba proteins and vCry1Ba proteins of the invention are further illustrated in Examples 2, 4, 5, 6 and 9.

In one embodiment, the present invention encompasses a nucleic acid that encodes an eCry1Ba protein of the invention 25 or encodes a vCry1Ba protein of the invention. In another embodiment, the nucleic acid comprises SEQ ID NO: 13.

The invention also encompasses a chimeric gene comprising a heterologous promoter sequence operatively linked to a nucleic acid that encodes an eCry1Ba protein or vCry1Ba 30 protein. In one embodiment, the heterologous promoter is selected from the group consisting of maize ubiquitin, cestrum virus (cmp), corn TrpA, rice actin, bacteriophage T3 gene 9 5' UTR, maize metallothionein (mtl), corn sucrose synthetase 1, corn alcohol dehydrogenase 1, corn light harvesting complex, corn heat shock protein, pea small subunit RuBP carboxylase, Ti plasmid opine synthase, Ti plaid nopaline synthase, petunia chalcone isomerase, bean glycine rich protein 1, Potato patatin, lectin, CaMV 35S, and the S-E9 small subunit RuBP carboxylase promoter

The present invention also encompasses recombinant vectors comprising the nucleic acid sequences of this invention. Such vectors include, without limitation, a plasmid, cosmid, phagemid, artificial chromosome, phage or viral vector. In such vectors, the nucleic acid sequences are preferably com- 45 prised in expression cassettes comprising regulatory elements for expression of the nucleic acid sequences in a host cell capable of expressing the nucleic acid sequences. Such regulatory elements usually comprise promoter and termination signals and preferably also comprise elements allowing 50 efficient translation of polypeptides encoded by the nucleic acid sequences of the invention. Vectors comprising the nucleic acid sequences are usually capable of replication in particular host cells, preferably as extrachromosomal molecules, and are therefore used to amplify the nucleic acid 55 sequences of the invention in the host cells. In one embodiment, host cells for such vectors are microorganisms, such as bacteria, including without limitation E. coli, Bacillus thuringiensis, Bacillus subtilis, Bacillus megaterium, Bacillus cereus, Agrobacterium or Pseudomonas. In another embodi- 60 ment, host cells for such recombinant vectors are endophytes or epiphytes. In another embodiment, the host cell for such vectors is a eukaryotic cell, such as a plant cell. Examples of such plant cells encompassed by the invention include, without limitation, sorghum, wheat, sunflower, tomato, potato, 65 cole crop, cotton, rice, soybean, sugar beet, sugarcane, tobacco, barley, oilseed rape or a corn cells.

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In another embodiment, such vectors are viral vectors and are used for replication of the nucleic acid sequences in particular host cells, e.g. insect cells or plant cells. Recombinant vectors are also used for transformation of the nucleic acid sequences of the invention into host cells, whereby the nucleic acid sequences are stably integrated into the DNA of such host cells. In one embodiment, such host cells are prokaryotic cells. In another embodiment, such host cells are eukaryotic cells, such as plant cells. In another embodiment, the plant cells are corn cells.

In one embodiment, the invention encompasses transgenic plants comprising a nucleic acid of the invention that encodes an eCry1Ba protein or vCry1Ba protein according to the invention. The eCry1Ba proteins or vCry1Ba proteins are useful for expressing in any transgenic plant where susceptible insect pests are a problem. Such transgenic plants include, without limitation, monocotyledonous plants and dicotyledous plants. In one embodiment, the monocotyledonous plants include corn, wheat, oat, rice, barley, sugar cane, sorghum, turf grass, and pasture grass plants. In another embodiment, the dicotyledonous plants include soybean and other legumes, cotton, sunflower, cole crops and other vegetables, sugar beet, tobacco and oilseed rape.

In another embodiment, the invention encompasses a progeny plant from any generation of a transgenic plant, wherein the progeny comprises a nucleic acid of the invention.

In yet another embodiment, the invention encompasses a propagule from any generation of a transgenic plant, wherein the propagule comprises a nucleic acid of the invention. In still another embodiment, the propagule of the invention is selected from the group consisting of a seed, a sette and a cutting.

In another embodiment, the invention encompasses a biological sample from a transgenic plant of the invention, wherein the biological sample comprises an eCry1Ba protein of the invention and the eCry1Ba protein is capable of controlling insect pests. Examples of such biological samples include without limitation any bi-product of corn that comprises protein such as corn meal or corn flour comprising the eCry1Ba protein, where the eCry1Ba protein continues to perform the insecticidal function it had in the transgenic corn plant from which the biological sample was derived.

The invention also encompasses an insecticidal composition comprising an eCry1B protein or vCry1Ba according to the invention and an acceptable agricultural carrier. In one embodiment, the agricultural carrier may be a liquid, a powder, or a transgenic plant, for example without limitation a corn plant or a sugar cane plant.

In another embodiment, the invention encompasses a method of producing an eCry1B protein or a vCry1Ba protein that is active against insects, comprising: (a) obtaining a host cell comprising a chimeric gene, which itself comprises a heterologous promoter sequence operatively linked to a nucleic acid of the invention; and (b) expressing the nucleic acid in the transgenic host cell, which results in at least one protein of the invention that is active against insects. In another embodiment, the insects are lepidopteran insects or coleopteran insects. In yet another embodiment, the lepidopteran insects are selected from the group consisting of European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean caterpillar.

In a further embodiment, the invention encompasses a method of producing an insect-resistant transgenic plant, comprising introducing an expression cassette comprising a nucleic acid of the invention into a plant thereby producing a transgenic plant, wherein the expression cassette causes the

expression of a protein of the invention in an amount that makes the plant resistant to insects. In another embodiment, the insects are lepidopteran or coleopteran insects. Such lepidopteran insects encompassed by the invention include without limitation European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean caterpillar.

In another embodiment, the invention encompasses a method of making an eCry1Ba protein comprising a) identifying a Cry1Ba protein having a domain I; b) substituting at 10 least one native amino acid at a location in domain I with at least one other amino acid; and c) obtaining the eCry1Ba protein so produced, wherein the eCry1Ba has improved solubility and/or insecticidal activity against at least European corn borer when compared to a native Cry1Ba protein. 15 In another embodiment, the Cry1Ba protein is Cry1Ba1 having GenBank accession number CAA29898, Cry1Ba2 (CAA65003), Cry1Ba3 (AAK63251), Cry1Ba4 Cry1Ba5 (AAK51084), (AB020894) or Cry1Ba6 (ABL60921). In still another embodiment, the native amino 20 acid in Cry1Ba is located in alpha-helix 4 or alpha-helix 5 of domain I. In still another embodiment the amino acid is at a position corresponding to position 150, 178, 189 or 199 of SEQ ID NO: 2. In yet another embodiment, the amino acid is at position 150, 178, 189 or 199 of SEQ ID NO: 5. In still 25 another embodiment, the amino acid in Cry1Ba is at a position corresponding to amino acids 2 and 150; or amino acids 2,150 and 178; or amino acids 2,150 and 189; or amino acids 2, 150 and 199, of SEQ ID NO: 5. In another embodiment, the amino acid is at positions 2 and 150; or positions 2, 150 and 30 178; or at positions 2, 150 and 189; or at positions 2, 150 and 199, of SEQ ID NO: 5.

In yet another embodiment, the invention encompasses a method of controlling insects, comprising delivering to the insects or contacting the insects with an effective amount of 35 an eCry1Ba protein or vCry1Ba protein of the invention. According to this embodiment, the insects are lepidopteran insects or coleopteran insects. Such lepidopteran insects include without limitation European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper 40 and velvet bean caterpillar. Preferably, the eCry1Ba protein or vCry1Ba protein is delivered to the insects orally. In another embodiment, the protein is delivered orally through a transgenic plant comprising a nucleic acid sequence that expresses an eCry1B protein or vCry1Ba protein of the invention.

The present invention further encompasses a method of controlling insects wherein a transgenic plant of the invention further comprises a second nucleic acid sequence or groups of nucleic acid sequences that encode a second pesticidal principle. In one embodiment, the second nucleic acid sequences are those that encode a Cry protein different than an eCry1Ba protein or vCry1Ba protein of the invention, those that encode a Vegetative Insecticidal Protein toxin, disclosed in U.S. Pat. Nos. 5,849,870 and 5,877,012, incorporated herein by reference, or those that encode a pathway for the production of a 55 non-proteinaceous principle. In another embodiment, the second nucleic acid sequence encodes a Vip3 protein. The skilled person will recognize that many different insecticidal principles may used in combination with an eCry1Ba or vCry1Ba protein of the invention.

In another embodiment, the invention encompasses a method of providing a grower with an improved means of controlling at least European corn borer comprising supplying or selling to the grower transgenic propagules comprising a nucleic acid that encodes an eCry1Ba protein having a 65 mutation at one or more amino acid positions in domain I, the eCry1Ba protein having improved solubility and/or insecti-

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cidal activity against at least European corn borer when compared to a native Cry1Ba protein. In another embodiment, the transgenic propagule is selected from the group consisting of a seed, a sette and a cutting.

In further embodiments, the nucleic acid sequences of the invention can be further modified by incorporation of random mutations in a technique known as in vitro recombination or DNA shuffling. This technique is described in Stemmer et al., Nature 370:389-391 (1994) and U.S. Pat. No. 5,605,793, which are incorporated herein by reference. Millions of mutant copies of a nucleic acid sequence are produced based on an original nucleic acid sequence of this invention and variants with improved properties, such as increased insecticidal activity, enhanced stability, or different specificity or ranges of target-insect pests are recovered. The method encompasses forming a mutagenized double-stranded polynucleic acid from a template double-stranded polynucleic acid comprising a nucleic acid sequence of this invention, wherein the template double-stranded polynucleic acid has been cleaved into double-stranded-random fragments of a desired size, and comprises the steps of adding to the resultant population of double-stranded random fragments one or more single or double-stranded oligonucleic acids, wherein said oligonucleic acids comprise an area of identity and an area of heterology to the double-stranded template polynucleic acid; denaturing the resultant mixture of double-stranded random fragments and oligonucleic acids into single-stranded fragments; incubating the resultant population of single-stranded fragments with a polymerase under conditions which result in the annealing of said single-stranded fragments at said areas of identity to form pairs of annealed fragments, said areas of identity being sufficient for one member of a pair to prime replication of the other, thereby forming a mutagenized double-stranded polynucleic acid; and repeating the second and third steps for at least two further cycles, wherein the resultant mixture in the second step of a further cycle includes the mutagenized double-stranded polynucleic acid from the third step of the previous cycle, and the further cycle forms a further mutagenized double-stranded polynucleic acid. In a preferred embodiment, the concentration of a single species of double-stranded random fragment in the population of double-stranded random fragments is less than 1% by weight of the total DNA. In a further embodiment, the template double-stranded polynucleic acid comprises at least about 100 species of polynucleic acids. In another preferred embodiment, the size of the double-stranded random fragments is from about 5 bp to 5 kb. In yet a further embodiment, the fourth step of the method comprises repeating the second and the third steps for at least 10 cycles.

Expression of the Nucleic Acid Sequences in Heterologous Microbial Hosts

As biological insect control agents, the insecticidal eCry1Ba proteins are produced by expression of the nucleic acid sequences in heterologous host cells capable of expressing the nucleic acid sequences. In a first embodiment, *B. thuringiensis* cells comprising modifications of a nucleic acid sequence of this invention are made. Such modifications encompass mutations or deletions of existing regulatory elements, thus leading to altered expression of the nucleic acid sequence, or the incorporation of new regulatory elements controlling the expression of the nucleic acid sequence. In another embodiment, additional copies of one or more of the nucleic acid sequences are added to *Bacillus thuringiensis* cells either by insertion into the chromosome or by introduction of extrachromosomally replicating molecules containing the nucleic acid sequences.

In another embodiment, at least one of the nucleic acid sequences of the invention is inserted into an appropriate expression cassette, comprising a promoter and termination signal. Expression of the nucleic acid sequence is constitutive, or an inducible promoter responding to various types of 5 stimuli to initiate transcription is used. In a preferred embodiment, the cell in which the protein is expressed is a microorganism, such as a virus, bacteria, or a fungus. In one embodiment, a virus, such as a baculovirus, contains a nucleic acid sequence of the invention in its genome and expresses large 10 amounts of the corresponding insecticidal eCry1Ba protein or vCry1Ba protein after infection of appropriate eukaryotic cells that are suitable for virus replication and expression of the nucleic acid sequence. The insecticidal protein thus produced is used as an insecticidal agent. Alternatively, baculovi- 15 ruses engineered to include the nucleic acid sequence are used to infect insects in vivo and kill them either by expression of the insecticidal protein or by a combination of viral infection and expression of the insecticidal protein.

Bacterial cells are also hosts for the expression of the 20 nucleic acid sequences of the invention. In a preferred embodiment, non-pathogenic symbiotic bacteria, which are able to live and replicate within plant tissues, so-called endophytes, or non-pathogenic symbiotic bacteria, which are capable of colonizing the phyllosphere or the rhizosphere, 25 so-called epiphytes, are used. Such bacteria include bacteria of the genera Agrobacterium, Alcaligenes, Azospirillum, Azotobacter, Bacillus, Clavibacter, Enterobacter, Erwinia, Flavobacter, Klebsiella, Pseudomonas, Rhizobium, Serratia, Streptomyces and Xanthomonas. Symbiotic fungi, such as 30 Trichoderma and Gliocladium are also possible hosts for expression of the inventive nucleic acid sequences for the same purpose.

Techniques for these genetic manipulations are specific for the different available hosts and are known in the art. For 35 example, the expression vectors pKK223-3 and pKK223-2 can be used to express heterologous genes in E. coli, either in transcriptional or translational fusion, behind the tac or trc promoter. For the expression of operons encoding multiple ORFs, the simplest procedure is to insert the operon into a 40 vector such as pKK223-3 in transcriptional fusion, allowing the cognate ribosome binding site of the heterologous genes to be used. Techniques for overexpression in gram-positive species such as Bacillus are also known in the art and can be used in the context of this invention (Quax et al. In:Industrial 45 Microorganisms:Basic and Applied Molecular Genetics, Eds. Baltz et al., American Society for Microbiology, Washington (1993)). Alternate systems for overexpression rely for example, on yeast vectors and include the use of *Pichia*, Saccharomyces and Kluyveromyces (Sreekrishna, In:Indus- 50 trial microorganisms:basic and applied molecular genetics, Baltz, Hegeman, and Skatrud eds., American Society for Microbiology, Washington (1993); Dequin & Bane, Biotechnology L2:173-177 (1994); van den Berg et al., Biotechnology 8:135-139 (1990)).

Plant Transformation

In one embodiment, at least one of the insecticidal eCry1B proteins or vCry1Ba proteins of the invention is expressed in a higher organism, e.g., a plant. In this case, transgenic plants expressing effective amounts of the eCry1Ba or vCry1Ba 60 proteins protect themselves from insect pests. When the insect starts feeding on such a transgenic plant, it also ingests the expressed eCry1Ba or vCry1Ba protein. This will deter the insect from further biting into the plant tissue or may even harm or kill the insect. A nucleic acid sequence of the present 65 invention is inserted into an expression cassette, which is then preferably stably integrated in the genome of said plant. In

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another embodiment, the nucleic acid sequence is included in a non-pathogenic self-replicating virus. Plants transformed in accordance with the present invention may be monocots or dicots and include, but are not limited to, corn, wheat, barley, rye, sweet potato, bean, pea, chicory, lettuce, cabbage, cauliflower, broccoli, turnip, radish, spinach, asparagus, onion, garlic, pepper, celery, squash, pumpkin, hemp, zucchini, apple, pear, quince, melon, plum, cherry, peach, nectarine, apricot, strawberry, grape, raspberry, blackberry, pineapple, avocado, papaya, mango, banana, soybean, tomato, sorghum, sugarcane, sugar beet, sunflower, rapeseed, clover, tobacco, carrot, cotton, alfalfa, rice, potato, eggplant, cucumber, *Arabidopsis*, and woody plants such as coniferous and deciduous trees.

Once a desired nucleic acid sequence has been transformed into a particular plant species, it may be propagated in that species or moved into other varieties of the same species, particularly including commercial varieties, using traditional breeding techniques.

A nucleic acid sequence of this invention may be expressed in transgenic plants, thus causing the biosynthesis of the corresponding eCry1Ba or vCry1Ba protein in the transgenic plants. In this way, transgenic plants with enhanced resistance to insects are generated. For their expression in transgenic plants, the nucleic acid sequences of the invention may require other modifications and optimization. Although in many cases genes from microbial organisms can be expressed in plants at high levels without modification, low expression in transgenic plants may result from microbial nucleic acid sequences having codons that are not preferred in plants. It is known in the art that all organisms have specific preferences for codon usage, and the codons of the nucleic acid sequences described in this invention can be changed to conform with plant preferences, while maintaining the amino acids encoded thereby. Furthermore, high expression in plants is best achieved from coding sequences that have at least about 35% GC content, preferably more than about 45%, more preferably more than about 50%, and most preferably more than about 60%. Microbial nucleic acid sequences that have low GC contents may express poorly in plants due to the existence of ATTTA motifs that may destabilize messages, and AATAAA motifs that may cause inappropriate polyadenylation. Although preferred gene sequences may be adequately expressed in both monocotyledonous and dicotyledonous plant species, sequences can be modified to account for the specific codon preferences and GC content preferences of monocotyledons or dicotyledons as these preferences have been shown to differ (Murray et al. Nucl. Acids Res. 17:477-498 (1989)). In addition, the nucleic acid sequences are screened for the existence of illegitimate splice sites that may cause message truncation. All changes required to be made within the nucleic acid sequences such as those described above are made using well known techniques of site directed mutagenesis, PCR, and synthetic gene construction using the methods described in the published patent applications EP 0 385 962 (to Monsanto), EP 0 359 472 (to Lubrizol, and WO 93/07278 (to Ciba-Geigy).

In one embodiment of the invention an eCry1Ba coding sequence is made according to the procedure disclosed in U.S. Pat. No. 5,625,136, herein incorporated by reference. In this procedure, maize preferred codons, i.e., the single codon that most frequently encodes that amino acid in maize, are used. The maize preferred codon for a particular amino acid might be derived, for example, from known gene sequences from maize. Maize codon usage for 28 genes from maize plants is found in Murray et al., Nucleic Acids Research 17:477-498 (1989), the disclosure of which is incorporated

herein by reference. A synthetic sequence made with maize optimized codons is set forth in SEQ ID NO: 13.

In this manner, the nucleic acid sequences can be optimized for expression in any plant. It is recognized that all or any part of the gene sequence may be optimized or synthetic. That is, 5 synthetic or partially optimized sequences may also be used.

For efficient initiation of translation, sequences adjacent to the initiating methionine may require modification. For example, they can be modified by the inclusion of sequences known to be effective in plants. Joshi has suggested an appropriate consensus for plants (NAR 15:6643-6653 (1987)) and Clonetech suggests a further consensus translation initiator (1993/1994 catalog, page 210). These consensuses are suitable for use with the nucleic acid sequences of this invention. The sequences are incorporated into constructions comprising the nucleic acid sequences, up to and including the ATG (whilst leaving the second amino acid unmodified), or alternatively up to and including the GTC subsequent to the ATG (with the possibility of modifying the second amino acid of the transgene).

Expression of the nucleic acid sequences in transgenic plants is driven by promoters that function in plants. The choice of promoter will vary depending on the temporal and spatial requirements for expression, and also depending on the target species. Thus, expression of the nucleic acid 25 sequences of this invention in leaves, in stalks or stems, in ears, in inflorescences (e.g. spikes, panicles, cobs, etc.), in roots, and/or seedlings is preferred. In many cases, however, protection against more than one type of insect pest is sought, and thus expression in multiple tissues is desirable. Although 30 many promoters from dicotyledons have been shown to be operational in monocotyledons and vice versa, ideally dicotyledonous promoters are selected for expression in dicotyledons, and monocotyledonous promoters for expression in monocotyledons. However, there is no restriction to the prov- 35 enance of selected promoters; it is sufficient that they are operational in driving the expression of the nucleic acid sequences in the desired cell.

Promoters that are expressed constitutively include promoters from genes encoding actin or ubiquitin and the CaMV 40 35S and 19S promoters. The nucleic acid sequences of this invention can also be expressed under the regulation of promoters that are chemically regulated. This enables the insecticidal eCry1Ba or variant Cry1Ba proteins to be synthesized only when the crop plants are treated with the inducing chemicals. Preferred technology for chemical induction of gene expression is detailed in the published application EP 0 332 104 (to Ciba-Geigy) and U.S. Pat. No. 5,614,395. A preferred promoter for chemical induction is the tobacco PR-1a promoter.

Another category of promoters is that which is wound inducible. Numerous promoters have been described which are expressed at wound sites and also at the sites of phytopathogen infection. Ideally, such a promoter should only be active locally at the sites of infection, and in this way the 55 insecticidal eCry1B or variant Cry1Ba proteins only accumulate in cells that need to synthesize the insecticidal eCry1Ba or variant Cry1Ba proteins to kill the invading insect pest. Promoters of this kind include those described by Stanford et al. Mol. Gen. Genet. 215:200-208 (1989), Xu et al. Plant 60 Molec. Biol. 22:573-588 (1993), Logemann et al. Plant Cell 1:151-158 (1989), Rohrmeier & Lehle, Plant Molec. Biol. 22:129-142 (1993), and Warner et al. Plant J. 3:191-201 (1993).

Tissue-specific or tissue-preferential promoters useful for 65 the expression of the eCry1Ba or variant Cry1Ba protein genes in plants, particularly corn, are those which direct

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expression in root, pith, leaf or pollen, particularly root. Such promoters, e.g. those isolated from PEPC or trpA, are disclosed in U.S. Pat. No. 5,625,136, or MTL, disclosed in U.S. Pat. No. 5,466,785. Both U.S. patents are herein incorporated by reference in their entirety.

Further preferred embodiments are transgenic plants expressing the nucleic acid sequences in a wound-inducible or pathogen infection-inducible manner.

In addition to promoters, a variety of transcriptional terminators are also available for use in chimeric gene construction using the eCry1Ba or variant Cry1Ba protein genes of the present invention. Transcriptional terminators are responsible for the termination of transcription beyond the transgene and its correct polyadenylation. Appropriate transcriptional terminators and those that are known to function in plants include the CaMV 35S terminator, the tml terminator, the nopaline synthase terminator, the pea rbcS E9 terminator and others known in the art. These can be used in both monocotyledons and dicotyledons. Any available terminator known to function in plants can be used in the context of this invention.

Numerous other sequences can be incorporated into expression cassettes described in this invention. These include sequences that have been shown to enhance expression such as intron sequences (e.g. from Adhl and bronzel) and viral leader sequences (e.g. from TMV, MCMV and AMV)

It may be preferable to target expression of the nucleic acid sequences of the invention to different cellular localizations in the plant. In some cases, localization in the cytosol may be desirable, whereas in other cases, localization in some subcellular organelle may be preferred. Subcellular localization of transgene-encoded enzymes is undertaken using techniques well known in the art. Typically, the DNA encoding the target peptide from a known organelle-targeted gene product is manipulated and fused upstream of the nucleic acid sequence. Many such target sequences are known for the chloroplast and their functioning in heterologous constructions has been shown. The expression of the nucleic acid sequences of the invention is also targeted to the endoplasmic reticulum or to the vacuoles of the host cells. Techniques to achieve this are well known in the art.

Vectors suitable for plant transformation are described elsewhere in this specification. For Agrobacterium-mediated transformation, binary vectors or vectors carrying at least one T-DNA border sequence are suitable, whereas for direct gene transfer any vector is suitable and linear DNA containing only the construction of interest may be preferred. In the case of direct gene transfer, transformation with a single DNA species or co-transformation can be used (Schocher et al. Biotechnology 4:1093-1096 (1986)). For both direct gene transfer and Agrobacterium-mediated transfer, transformation is usually (but not necessarily) undertaken with a selectable marker that may provide resistance to an antibiotic (kanamycin, hygromycin or methotrexate) or a herbicide (basta). Plant transformation vectors comprising the eCry1Ba or variant Cry1Ba protein genes of the present invention may also comprise genes, for example, phosphomannose isomerase (pmi), which provides for positive selection of the transgenic plants as disclosed in U.S. Pat. Nos. 5,767,378 and 5,994,629, herein incorporated by reference, or phosphinotricin acetyltransferase (pat), which provides tolerance to the herbicide phosphinotricin (glufosinate). The choice of selectable marker is not, however, critical to the invention.

In another embodiment, a nucleic acid sequence encoding an eCry1Ba or vCry1Ba protein of the invention is directly transformed into the plastid genome. A major advantage of plastid transformation is that plastids are generally capable of

expressing bacterial genes without substantial codon optimization, and plastids are capable of expressing multiple open reading frames under control of a single promoter. Plastid transformation technology is extensively described in U.S. Pat. Nos. 5,451,513, 5,545,817, and 5,545,818, in PCT appli- 5 cation no. WO 95/16783, and in McBride et al. (1994) Proc. Nati. Acad. Sci. USA 91, 7301-7305. The basic technique for chloroplast transformation involves introducing regions of cloned plastid DNA flanking a selectable marker together with the gene of interest into a suitable target tissue, e.g., 10 using biolistics or protoplast transformation (e.g., calcium chloride or PEG mediated transformation). The 1 to 1.5 kb flanking regions, termed targeting sequences, facilitate homologous recombination with the plastid genome and thus allow the replacement or modification of specific regions of 15 the plastome. Initially, point mutations in the chloroplast 16S rRNA and rps12 genes conferring resistance to spectinomycin and/or streptomycin are utilized as selectable markers for transformation (Svab, Z., Hajdukiewicz, P., and Maliga, P. (1990) Proc. Nati. Acad. Sci. USA 87, 8526-8530; Staub, J. 20 M., and Maliga, P. (1992) Plant Cell 4, 39-45). This resulted in stable homoplasmic transformants at a frequency of approximately one per 100 bombardments of target leaves. The presence of cloning sites between these markers allowed creation of a plastid targeting vector for introduction of for- 25 eign genes (Staub, J. M., and Maliga, P. (1993) EMBO J. 12, 601-606). Substantial increases in transformation frequency are obtained by replacement of the recessive rRNA or r-protein antibiotic resistance genes with a dominant selectable marker, the bacterial aadA gene encoding the spectinomycincletoxifying enzyme aminoglycoside-3'-adenyltransf erase (Svab, Z., and Maliga, P. (1993) Proc. Natl. Acad. Sci. USA 90, 913-917). Previously, this marker had been used successfully for high-frequency transformation of the plastid genome of the green alga Chlamydomonas reinhardtii (Goldschmidt- 35 Clermont, M. (1991) Nucl. Acids Res. 19:4083-4089). Other selectable markers useful for plastid transformation are known in the art and encompassed within the scope of the invention. Typically, approximately 15-20 cell division cycles following transformation are required to reach a 40 homoplastidic state. Plastid expression, in which genes are inserted by homologous recombination into all of the several thousand copies of the circular plastid genome present in each plant cell, takes advantage of the enormous copy number advantage over nuclear-expressed genes to permit expression 45 levels that can readily exceed 10% of the total soluble plant protein. In a preferred embodiment, a nucleic acid sequence of the present invention is inserted into a plastid-targeting vector and transformed into the plastid genome of a desired plant host. Plants homoplastic for plastid genomes containing 50 a nucleic acid sequence of the present invention are obtained, and are preferentially capable of high expression of the nucleic acid sequence.

Combinations of Insect Control Principles

The eCry1Ba or vCry1Ba proteins of the invention can be 55 used in combination with other Bt Cry proteins or other pesticidal principles to increase pest target range. Furthermore, the use of the eCry1Ba or vCry1Ba proteins of the invention in combination with other Bt Cry proteins or other pesticidal principles of a distinct nature has particular utility 60 for the prevention and/or management of insect resistance. Other insecticidal principles include, for example, lectins,  $\alpha$ -amylase, peroxidase and cholesterol oxidase. Vegetative Insecticidal Protein genes, such as vip1A(a) and vip2A(a) or vip3, are also useful in the present invention. In one embodiment, an eCry1Ba protein designated eCry1Ba-T2AY150KL189S (SEQ ID NO: 9) is combined with a Vip3A

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protein in a transgenic plant. The transgenic plant exhibits the combined spectrum of insecticidal activity associated with both the eCry1Ba and Vip3. In yet another embodiment, the transgenic plant is a corn plant or a sugar cane plant.

This co-expression of more than one insecticidal principle in the same transgenic plant can be achieved by genetically engineering a plant to contain and express all the genes necessary in a so called molecular stack. Alternatively, a plant, Parent 1, can be genetically engineered for the expression of genes of the present invention. A second plant, Parent 2, can be genetically engineered for the expression of a supplemental insect control principle. By crossing Parent 1 with Parent 2, progeny plants are obtained which express all the genes introduced into Parents 1 and 2. For example without limitation, Parent 1 may contain an eCry1Ba coding sequence and Parent 2 may contain a Vip3A coding sequence. Some progeny of a Parent 1×Parent 2 cross will contain both the eCry1Ba coding sequence and the Vip3A coding sequence.

Transgenic seed of the present invention can also be treated with an insecticidal seed coating as described in U.S. Pat. Nos. 5,849,320 and 5,876,739, herein incorporated by reference. Where both the insecticidal seed coating and the transgenic seed of the invention are active against the same target insect, the combination is useful (i) in a method for enhancing activity of an eCry1Ba protein of the invention against the target insect and (ii) in a method for preventing development of resistance to an eCry1Ba protein of the invention by providing a second mechanism of action against the target insect. Thus, the invention provides a method of enhancing activity against or preventing development of resistance in a target insect, for example corn rootworm, comprising applying an insecticidal seed coating to a transgenic seed comprising one or more eCry1Ba proteins of the invention. Such chemical treatments may include insecticides, fungicides or nematicides. Examples of such insecticides include, without limitation, dinotefuran, such as thiamethoxam, imidacloprid, acetamiprid, nitenpyram, nidinotefuran, chlorfenapyr, tebufenpyrad, tebufenozide, methoxyfenozide, halofenozide, triazamate, avermectin, spinosad, fiprinol, acephate, fenamichlorpyrifos, diazinon, chlorpyrifon-methyl, malathion, carbaryl, aldicarb, carbofuran, thiodicarb, and oxamyl. Even where the insecticidal seed coating is active against a different insect, the insecticidal seed coating is useful to expand the range of insect control, for example by adding an insecticidal seed coating that has activity against lepidopteran insects to the transgenic seed of the invention, which has activity against coleopteran insects, the coated transgenic seed produced controls both lepidopteran and coleopteran insect pests.

#### **EXAMPLES**

The invention will be further described by reference to the following detailed examples. These examples are provided for the purposes of illustration only, and are not intended to be limiting unless otherwise specified. Standard recombinant DNA and molecular cloning techniques used here are well known in the art and are described by J. Sambrook, et al., Molecular Cloning: A Laboratory Manual, 3d Ed., Cold Spring Harbor, N.Y.: Cold Spring Harbor Laboratory Press (2001); by T. J. Silhavy, M. L. Berman, and L. W. Enquist, Experiments with Gene Fusions, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1984) and by Ausubel, F. M. et al., Current Protocols in Molecular Biology, New York, John Wiley and Sons Inc., (1988), Reiter, et al., Methods in

*Arabidopsis Research*, World Scientific Press (1992), and Schultz et al., *Plant Molecular Biology Manual*, Kluwer Academic Publishers (1998).

#### Example 1

#### Use of PCR to Mutate Cry1Ba Coding Sequences

Polymerase Chain Reaction (PCR) is a repetitive, enzymatic, primed synthesis of a nucleic acid sequence. This 10 procedure is well known and commonly used by those skilled in this art (See Mullis, U.S. Pat. Nos. 4,683,195, 4,683,202, and 4,800,159; Saiki, Randall K., Stephen Scharf, Fred Faloona, Kary B. Mullis, Glenn T. Horn, Henry A. Erlich, Norman Arnheim [1985] "Enzymatic Amplification of 15  $\beta$ -Globin Genomic Sequences and Restriction Site Analysis for Diagnosis of Sickle Cell Anemia," Science 230:1350-1354.). PCR is based on the enzymatic amplification of a DNA fragment of interest that is flanked by two oligonucleic acid primers that hybridize to opposite strands of the target 20 sequence. The primers are oriented with the 3' ends pointing

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towards each other. Repeated cycles of heat denaturation of the template, annealing of the primers to their complementary sequences, and extension of the annealed primers with a DNA polymerase result in the amplification of the segment defined by the 5' ends of the PCR primers. Since the extension product of each primer can serve as a template for the other primer, each cycle essentially doubles the amount of DNA fragment produced in the previous cycle. This results in the exponential accumulation of the specific target fragment, up to several million-fold in a few hours. By using a thermostable DNA polymerase such as Taq polymerase, which is isolated from the thermophilic bacterium *Thermus aquaticus*, the amplification process can be completely automated.

The mutant Cry1Ba coding sequences described in the following examples were constructed using a QuickChange Site-Directed Mutagenesis Kit (Stratagene, La Jolla, Calif.) according to the manufacturer's instructions and various combinations of the exemplified primers shown in Table 1. The skilled person will recognize based on the present application that other primer pairs can be used to mutate any Cry1Ba coding sequence.

TABLE 1

	Primers used to make muated coding sequence encoding	eCrylBa proteins.
Primer Name	Primer Sequence	SEQ ID NO:
YG152	5'-agaagtgttcttnnsacccaatatatagctttagaacttg-3'	SEQ ID NO: 14
YG153	5'-tatatattgggtsnnaagaacacttctcgttcttgcatc-3'	SEQ ID NO: 15
YG154	5'-agaagtgttcttaagacccaatatatagctttagaacttg-3'	SEQ ID NO: 16
YG155	5'-tatatattgggtcttaagaacacttctcgttcttgcatc-3'	SEQ ID NO: 17
YG156	5'-agaagtgttctttggacccaatatatagctttagaacttg-3'	SEQ ID NO: 18
YG157	5'-tatatattgggtccaaagaacacttctcgttcttgcatc-3'	SEQ ID NO: 19
YG160	5'-atatgtttaaacatgacttcaaataggaaaaatgagaatgaa-3'	SEQ ID NO: 20
YG161	5'-atatgtttaaacatggatctattaccagatgctcgtattg-3'	SEQ ID NO: 21
YG162	5'-atatggcgcgcctatctttctaaatcatattctgcttcgaagg-3'	SEQ ID NO: 22
YG163	5'-aattccatggcgtcaaataggaaaaatgagaatgaaattataaatgc-3'	SEQ ID NO: 23
YG164	5'-aattccatggatctattaccagatgctcgtattg-3'	SEQ ID NO: 24
YG165	5'-aattccatggaggatagcttgtgtatagccgagg-3'	SEQ ID NO: 25
YG166	5'-aattgagctcttatctttctaaatcatattctgcttcgaagg-3'	SEQ ID NO: 26
YG171	5'-agttttctttggggtgaattatggccccgc-3'	SEQ ID NO: 27
YG172	5'-taattcaccccaaagaaaactataaaaactagc-3'	SEQ ID NO: 28
YG175	5'-caatatatagatttagaacttgattttcttaatg-3'	SEQ ID NO: 29
YG176	5'-aagttctaaatctatatattgggtataaagaac-3'	SEQ ID NO: 30
YG179	5'-ttacacctatccttattgagagatgcctctc-3'	SEQ ID NO: 31
YG180	5'-tctcaataaggataggtgtaaatttgcagcttg-3'	SEQ ID NO: 32
YG183	5'-agaacgagaagtgaacttaagacccaatatatagc-3'	SEQ ID NO: 33
YG184	5'-acccaatatatagatttagaacttgattttcttaatgcg-3'	SEQ ID NO: 34
YG186	5'-gaagttccattattgccggtatatgctcaagctgc-3'	SEQ ID NO: 35
YG188	5'-tttcttaataagatgccgcttttcgcaattagaaacc-3'	SEQ ID NO: 36
YG189	$\verb  5'-aagcggcatcttattaagaaaatcaagttctaaagctatatattggg-3'    $	SEQ ID NO: 37

TABLE 1-continued

	Primers used to make muated coding sequence encodi	nq eCrylBa	proteins.
Primer Name	Primer Sequence	SEQ ID	NO:
YG190	5'-ctttttggtaaggaatttgggcttacatcgcagg-3'	SEQ ID	NO: 38
YG191	5'-cccaaattccttaccaaaaagagaggcatctctcaat-3'	SEQ ID	NO: 39
YG192	5'-ccattattgagcgtatatgctcaagctgcaaatttacacc-3'	SEQ ID	NO: 40
YG193	5'-agcatatacgctcaataatggaacttcttggtttctaattgcg-3'	SEQ ID	NO: 41

#### Example 2

#### Determining Toxicity of Cry1Ba Mutants

Activity of mutant Cry1Ba proteins (described below) against insect pests, including European corn borer (Ostrinia 20 nubilalis), sugarcane borer (Diatraea saccharalis), southwestern corn borer (Diatraea grandiosella), corn earworm (Helicoverpa zea), soy bean looper (Pseudoplusia includens), velvet bean caterpillar (Anticarsia gemmatalis), and Colorado potato beetle (Leptinotarsa decemlineata) is 25 determined by a surface contamination method. Briefly, artificial diet for a particular species is poured into 24 well tissue culture plates or small petri dishes. Each well has a surface area of approximately 2 cm<sup>2</sup>. Liquids comprising the mutant Cry1Ba proteins are applied to the surface of the diet in each 30 well. After the liquid is absorbed and dried, test larvae are placed in each well and the then the plate is sealed. Activity of an engineered Cry1Ba protein is compared to a native or wild-type Cry1Ba and recorded as percent mortality or relative activity.

#### Example 3

#### Mutations at Position 150 in Full-Length Cry1Ba

Since the six native Cry1Ba proteins vary by only one amino acid in alpha-helix 4, e.g. 4/6 have a tyrosine (Y150) and 2/6 have a histidine (H150) (See FIG. 1), the initial mutagenesis analysis investigated the impact of amino acid position 150 in alpha-helix 4 on the insecticidal activity of 45 full-length Cry1Ba.

A native full-length cry1Ba coding sequence (SEQ ID NO: 1) was cloned into a Bt/E. coli pUC18-derived shuttled vector under the control of a Cry1Ac promoter. Using this full-length coding sequence as a template, mutant Cry1Ba proteins were generated by randomly substituting the tyrosine (Tyr) at position 150 with different amino acids using the QuickChange Site-Directed Mutagenesis Kit (Stratagene, La Jolla, Calif.) according to the manufacturer's instructions and primers YG152-YG157 of Table 1. All mutant Cry1Ba proteins were tested against ECB using the method described in Example 2.

The data shown in Table 2 demonstrate that position 150 in the full-length Cry1Ba protein plays an important role in modulating at least ECB toxicity. Several of the mutations 60 reduced the ECB specific activity compared to the native Cry1Ba. Cry1Ab, a protein with high specific activity against ECB, has an arginine (Arg) at the position corresponding to Y150 of the Cry1Ba sequence (position 131 of the Cry1Ab sequence; See FIG. 2A). Interestingly, the Y150R Cry1Ba 65 mutant had only half the activity of the native Cry1Ba protein used in the experiment. Mutations that maintained or slightly

15 increased the activity over native Cry1Ba included Y150K, Y150F, Y150W and Y150P. The mutant Cry1Ba proteins that had lower activity compared to native Cry1Ba were designated as variant Cry1Ba proteins.

TABLE 2

Mutant Designation	Amino Acid at Position 150	Relative ECB Mortality
M4	K	++++
M18	F	+++
M7	$\mathbf{W}$	+++
M26	P	+++
M12	T	++
M14	L	++
M15	A	++
M23	V	++
M28	S	++
M38	R	+
M9	G	+
M24	D	+/-
Native Cry1Ba	Y	+++
Empty Vector	_	_
(Control)		

#### Example 4

## Toxicity of Truncated Cry1Ba Compared to Full-Length Cry1Ba

Based on what is known in the art, it is not clear what exact proteolytic sites in Cry1Ba are targeted by insect gut proteases. Therefore, the sequence of the active toxin is unclear. For this example, cleavage sites for Cry1Ba protoxin were predicted based on a sequence alignment with Cry1Ab (See FIG. 2), whose cleavage sites have been reported. Using this information, vectors that express truncated versions of Cry1Ba were constructed.

A truncated cry1Ba fragment was PCR cloned into vector pCIB5634 or pET28a using the full-length native cry1Ba coding sequence (SEQ ID NO: 1) as the template and primers YG160 and YG162 or YG163 and YG166, respectively. The resulting PCR fragment encodes a truncated protein comprising amino acids 1-647 of SEQ ID NO: 2. However, during the initial cloning of the truncated cry1Ba coding sequence into the vectors, a mutation was introduced whereby the threonine at position 2 (T2) was replaced with an alanine (Ala; A) (T2A mutation). This T2A mutation was determined to have no negative impact on insecticidal activity compared to the native Cry1Ba and therefore was used in all subsequent mutation experiments. This T2A mutant was designated the T25 wild-type Cry1Ba.

60

65

30
TABLE 4-continued

Another truncated cry1Ba fragment was PCR cloned into vector pCIB5634 or pET28a using the T25 coding sequence as a template and primers YG161 and YG162 or YG164 and YG166, respectively. The resulting PCR fragment encodes an N-terminal and C-terminal truncated protein (SEQ ID NO: 542) comprising amino acids 22-647 of SEQ ID NO: 2 and was designated T7.

Western blot results demonstrate that the T25 truncated Cry1Ba (comprising amino acids 1-647) in both the pCIB5634 and pET28a vectors was more stable than the T7 truncated Cry1Ba toxin in either vector. Results of the bioassay (Table 3) showed that the T25 wild-type toxin was 15× more active than the T7 construct and 3× more active than the full-length Cry1Ba protein. Therefore, further Cry1Ba 15 mutants were constructed using the wild-type T25 truncated Cry1Ba

TABLE 3

Activity of truncated vs. full-length Cry1Ba against ECB.						
Clone	Amino acids	Activity Relative to Full-Length Cry1Ba				
T25	1-647	3.0				
T7	22-647	0.2				
FL-Cry1Ba	1-1228	1.0				
Vector Control		0.0				

Example 5

#### Effects of Mutating Y150 in Truncated Cry1Ba

The mutations at amino acid position 150 that did not decrease insecticidal activity, including Y150K, Y150F, Y150W and Y150P, of the full-length Cry1Ba protein were tested in the T25 truncated Cry1Ba toxin. The Y150K, Y150F, Y150W and Y150P mutations were made as described above 40 using the YG152-YG157 primers.

Making these mutations in the truncated T25 protein lead to different results than in the full-length Cry1Ba protein. For example, the Y150P mutation in the T25 truncated toxin completely knocked out ECB activity. However, the same mutation in the full-length Cry1Ba had no negative impact on ECB activity (See Example 3). Surprisingly, all the mutations except the Y150K mutation reduced the T25 ECB activity to some degree (Table 4). All known native Cry1Ba proteins have either a histidine (H) or a tyrosine (Y) at position 150. The Y150K mutation substantially altered the biological properties of the eCry1Ba-Y150K mutant compared to both the "H150-type" native Cry1Ba and the "Y150-type" native Cry1Ba. The Y150K mutant was 3 times more active than a Cry1Ba protein with a histidine (His) at position 150.

TABLE 4

Activity of truncated Y150X mutants.					
Mutant Designation	Amino Acid at Position 150	ECB Activity Relative to T25			
TM9	P	0.00			
TM5	F	0.25			
TM15	H	0.58			
TM27	W	0.75			
TM2	K	1.60			

 Activity of truncated Y150X mutants.

 Amino Acid at Mutant Designation
 ECB Activity Position 150
 Relative to T25

 T25 (wt)
 Y
 1.00

 Empty Vector (Control)
 —
 0.00

Each of the mutant T25 Cry1Ba proteins was tested for its solubility properties. Solubility of the proteins correlated with insecticidal activity. For example, the eCry1Ba-Y150K protein was more soluble than wild-type T25-Cry1Ba and any of the other mutant proteins. Therefore, these data demonstrate that changing the amino acid at position 150 has a dramatic impact on the solubility and insecticidal activity of a truncated Cry1Ba protein. For example, mutating the tyrosine (Tyr) at position 150 to lysine (Lys) substantially increases the solubility and specific activity of the truncated Cry1Ba toxin against ECB compared to the wild-type truncated Cry1Ba toxin (T25). The eCry1Ba-Y150K protein (TM2) was used for further mutational analysis experiments.

#### Example 6

#### Construction and Testing of Additional eCry1Ba Mutants

Cry1Ab has a high specific activity against ECB. Therefore, a sequence alignment was done between a Cry1Ab and the T25-Cry1Ba protein to help identify key amino acid positions in alpha-helix 4 or 5 that may be important to Cry1Ba activity or solubility. The sequence alignment between Cry1Ab and Cry1Ba is shown in FIG. 2. A comparison of the structural features of Cry1Ab and Cry1Ba is shown in Table 8 in Example 10 below. Further mutational analysis was carried out on the identified key amino acid positions to determine whether mutations in addition to the Y150K mutation would further increase the specific activity of this eCry1Ba protein. The TM2 coding sequence (SEQ ID NO: 4) was used as a template for further site-directed mutagenesis. The mutations were made as described above using the YG171-YG193 primers listed in Table 1.

Eleven mutants were tested for activity against European corn borer. Table 5 shows the results of the bioassays. Of the 11 mutants tested, two mutations, L189S and S199K, increased the specific activity of the TM2-Y150K mutant against ECB, which was at least a 3× increase in specific activity over the wild-type Cry1Ba (T25). These were designated as engineered Cry1Ba (eCry1Ba) proteins. Two mutations, V81W and M178S/A155S, had the same activity as TM2, and two mutants, M178P and R1705, had less activity than TM2. These mutants were categorized as variant Cry1Ba proteins (vCry1Ba). Four mutations, V148E/A155D, A155K, A163K and A163K/L188P knocked out activity completely, indicating that these positions are critical for at least ECB activity.

TABLE 5

Activity of TM2-Cry1Ba mutants compared to wild-type Cry1Ba.							
Clone	Mutations	Relative Activity	SEQ ID NO:				
T25	T2A (wt)	1.0	SEQ ID NO: 5				
TM2	Y150K	2.0	SEQ ID NO: 7				
TM21	Y150K/V81W	1.0	SEO ID NO: 11				

Activity of TM2-Cry1Ba mutants compared to wild-type Cry1Ba Mutations Relative Activity SEO ID NO: Y150K/V148E/A155D Y150K/L189S 3.0 SEQ ID NO: 9 Y150K/M178S SEO ID NO: 8 SEQ ID NO: 12 Y150K/M178S/A155S 1.0 Y150K/M178P 0.5 SEQ ID NO: 43 Y150K/R170S 0.5 SEQ ID NO: 44 Y150K/A155K 0.0

0.0

0.0

3.0

SEQ ID NO: 10

#### Spectrum of eCry1Ba Protein

Y150K/A163K

Y150K/S199K

Y150K/A163K/L188P

Clone

TM60

TM33

TM88

TM90

TM69

TM61

TM70

TM78

TM82

TM83

The TM33 mutant (eCry1Ba-T2AY150KL189S) was tested against several other lepidopteran insects, including, sugarcane borer (SCB; *Diatraea saccharalis*), southwestern corn borer (SWCB; *Diatraea grandiosella*), corn earworm (CEW, *Helicoverpa zea*), velvet bean caterpillar (VBC; *Anticarsia gemmatalis*), and soybean looper (SBL, *Pseudoplusia includens*, now named *Chrysodeixis includens*), using a surface treated artificial diet bioassays. Larval mortality was assessed after approximately 4-6 days depending on the 25 insect species tested.

Native Cry1Ba has been reported to be active against sugarcane borer, southwest corn borer and soybean looper and to have no activity against corn earworm. In addition, some reports have suggested that Bt strains comprising a Cry1B-type protein have activity against velvet bean caterpillar (Bobrowski et al. 2001. Brazil. J. Microbol. 32:105-109), but it is not clear from this report whether this activity is due to a Cry1Ba protein or to some other protein expressed in the Bt strain tested. Other reports (For example, Monnerat et al. 2007. Biological Control 41:291-295) demonstrate that Cry1B present in Bt strains contributes little to toxicity of such strains to VBC larvae.

Results of the bioassay of the eCry1Ba-T2AY150KL189S mutant showed that this protein, like the native Cry1Ba protein, is active against sugarcane borer, southwestern corn borer and soybean looper. Unlike the native Cry1Ba protein, the eCry1Ba protein was very active against velvet bean caterpillar. Surprisingly, the eCry1Ba protein also had some 45 activity against corn earworm, an insect for which native Cry1Ba has no activity. The activity of eCry1Ba protein against velvet bean caterpillar and corn earworm is another indication that eCry1Ba is substantially different from native Cry1Ba protein.

Since native Cry1Ba is known to be active against both lepidopteran and coleopteran insects, the TM33 eCry1Ba protein was tested against the coleopteran insect, Colorado potato beetle (CPB; Leptinotarsa decemlineata). Bioassays were carried out using neonate CPB larvae and a standard 55 artificial diet assay as described in Example 2 above. As was already known in the art, the native Cry1Ba protein was active against CPB. The wild-type Cry1Ba mutant, T25, was also active. Surprisingly, the T33 eCry1Ba protein was not active against CPB. Therefore, although the mutations in T33 60 increase the specific activity against at least European corn borer, these mutations knocked out the activity against the coleopteran insect, Colorado potato beetle, which is yet another indication that the properties of eCry1Ba proteins are substantially different than native Cry1Ba and wild-type Cry1Ba proteins. Using this approach the skilled person will recognize that mutation of amino acids in domain I, particu32

larly alpha-helix 4 and alpha-helix 5, of a Cry1Ba provides a method to change the spectrum of activity of Cry1Ba.

The mutants described above were tested for differences in solubility properties using standard methods known in the art. Briefly, cell pellets from induced E. coli cultures expressing Cry1Ba mutants and wild type Cry1Ba were treated in Bug-Buster<sup>TM</sup> protein extraction reagent (Novagen, Inc) with protease inhibitors and lysonase according to the manufacturer's instructions. Cell lysates and soluble fractions after centrifugation of cell lysates were analyzed on SDS-PAGE and western blot using rabbit-anti-Cry1Ba antibody, and Cry1Ba protein on western blot were quantified by AlphaImager (Cell Biosciences). Although the mutant Cry1Ba and the wild-type T25 showed similar level of protein expression in cell lysates, the amount of protein present in soluble fractions was surprisingly quite different between mutants and wild type. To compare solubility, Cry1Ba mutant proteins present in soluble fractions were normalized over wild type Cry1Ba. Results in Table 6 demonstrate that eCry1Ba mutants had in the range of 1.5 to 2.1 times more soluble eCry1Ba protein than the wild-type T25 Cry1Ba protein in the same amount of liquid under the same environmental conditions. "SP" in Table 6 means soluble protein; and "TP" means total protein.

TABLE 6

	Solubility comparison of eCry1Ba proteins and wild-type T25 Cry1Ba protein.						
)	Clone	Mutations	Percent SP/TP	Fold Increase over Wild-type (T25)			
	T25 (Wild-type)	T2A	52	1.0			
	TM33	T2A, Y150K, L189S	76	1.5			
	TM2	T2A, Y150K	86	1.7			
	TM83	T2A, Y150K, S199K	107	2.1			

#### Example 7

#### Maize-Optimized ecry1B Gene Construction

A maize optimized nucleotide sequence (mocry1Ba-TM33) that encodes the TM33 Cry1Ba mutant (eCry1Ba-T2A:Y150K:L189S) protein was generated as described in U.S. Pat. No. 6,051,760, herein incorporated by reference. The mocry1Ba-TM33 coding sequence is set forth in SEQ ID NO: 13. The eCry1Ba-T2A:Y150K:L189S amino acid sequence is set forth in SED ID NO: 9.

#### Example 8

#### Transgenic Maize and Sugarcane Expressing eCry1Ba Protein

Two plant transformation vectors are constructed for introduction of the mocry1Ba-TM33 coding sequence into maize: (a) a first vector (18320) comprising two expression cassettes, a first expression cassette comprising a maize ubiquitin promoter (ZmUbiInt) (Christensen et al. 1992 PMB 18: 675) operably linked to the TM33 coding sequence further operably linked to a nopaline synthase 3' end transcription termination and polyadenylation sequence, designated as ZmUbi: mocry1Ba-TM33:NOS, and a second expression cassette comprising a 35S:pat:NOS, and (b) a second vector (18319) comprising two expression cassettes, a first expression cassette comprising a MTL promoter sequence (U.S. Pat. No. 6,018,099) operably linked to the TM33 coding sequence further operably linked to a nopaline synthase 3' end tran-

Example 9

scription termination and polyadenylation sequence, designated as MTL:mocry1Ba-TM33:NOS, and a second expression cassette comprising 35S:pat:NOS. All vectors in this example comprise the pat gene encoding a phosphinotricin acetyltransferase (PAT), which confers tolerance to the herbicide phosphinotricin for selection of transgenic events.

Both vectors are individually transformed into maize. *Agrobacterium* transformation of immature maize embryos is performed essentially as described in Negrotto et al., 2000, Plant Cell Reports 19: 798-803. For this example, all media constituents are essentially as described in Negrotto et al., supra. However, various media constituents known in the art may be substituted.

Briefly, *Agrobacterium* strain LBA4404 (pSB1) containing a plant transformation plasmid is grown on YEP (yeast extract (5 g/L), peptone (10 g/L), NaCl (5 g/L), 15 g/l agar, pH 6.8) solid medium for 2-4 days at 28° C. Approximately  $0.8\times10^9$  *Agrobacterium* are suspended in LS-inf media supplemented with 100  $\mu$ M As (Negrotto et al., supra). Bacteria are pre-induced in this medium for 30-60 minutes.

Immature embryos from a suitable genotype are excised from 8-12 day old ears into liquid LS-inf+100  $\mu M$  As. Embryos are rinsed once with fresh infection medium. Agrobacterium solution is then added and embryos are vortexed 25 for 30 seconds and allowed to settle with the bacteria for 5 minutes. The embryos are then transferred scutellum side up to LSAs medium and cultured in the dark for two to three days. Subsequently, between 20 and 25 embryos per petri plate are transferred to LSDc medium supplemented with 30 cefotaxime (250 mg/l) and silver nitrate (1.6 mg/l) and cultured in the dark for 28° C. for 10 days.

Immature embryos, producing embryogenic callus are transferred to LSD1M0.5S medium. The cultures are selected on this medium for about 6 weeks with a subculture step at 35 about 3 weeks. Surviving calli are transferred to Reg1 medium supplemented with mannose. Following culturing in the light (16 hour light/8 hour dark regiment), green tissues are then transferred to Reg2 medium without growth regulators and incubated for about 1-2 weeks. Plantlets are transferred to Magenta GA-7 boxes (Magenta Corp, Chicago Ill.) containing Reg3 medium and grown in the light. After about 2-3 weeks, plants are tested for the presence of the pat gene and the mocry1Ba-TM33 coding sequence. Positive plants from the PCR assay are transferred to the greenhouse and 45 tested for resistance to at least European corn borer. Sugarcane Transformation

A plant transformation vector (72581) comprising two expression cassettes was constructed for introduction of the mocry1Ba-TM33 coding sequence into sugarcane. The first 50 expression cassette comprises a maize Ubi361 promoter (PCT/US10/37683) operably linked to the TM33 coding sequence further operably linked to a maize Ubi361 3' end transcription termination and polyadenylation sequence (PCT/US10/37683), prZmUbi361-3: 55 designated as mocry1Ba-TM33:tZmUbi361. The second expression cassette comprises a maize ubiquitin promoter (ZmUbiInt) (Christensen et al. 1992 PMB 18: 675) operably linked to a pmi coding sequence further operably linked to a nopaline synthase (nos) 3' end transcription termination and polyadenylation sequence. The pmi coding sequence encodes phosphomannose isomerase (PMI), which enables transgenic sugarcane to utilize mannose and functions as a selectable marker for transformation. The 72581 vector was transformed into sugarcane using Agrobacterium transformation. Transgenic 65 sugarcane plants were tested against neonate sugarcane borer as described above.

Insecticidal Activity of Transgenic Maize and Sugarcane Plants

Plants were sampled as they are being transplanted from Magenta GA-7 boxes into soil. Sampling consisted of cutting two small pieces of leaf (ca. 2-4 cm long) and placing each in a small petri dish or multi-well plates. Negative controls were either transgenic plants that were PCR negative for the mocry 1Ba-TM33 gene from the same experiment, or from non-transgenic plants (of a similar size to test plants) that are being grown in a greenhouse or phytotron.

Leaf samples from each plant were inoculated with a suitable target insect pest by placing approximately 10 first instar larvae onto each leaf piece. Petri dishes or multi-well plates were then tightly sealed.

0.8×10<sup>9</sup> *Agrobacterium* are suspended in LS-inf media supplemented with 100 μM As (Negrotto et al., supra). Bacteria are pre-induced in this medium for 30-60 minutes.

Immature embryos from a suitable genotype are excised from 8-12 day old ears into liquid LS-inf+100 μM As.

Embryos are rinsed once with fresh infection medium. *Agro*-

Results shown in Table 7 indicate that transgenic maize plants comprising the moTM33 gene and expressing the eCry1Ba-T2A:Y150K:L189S mutant protein, are insecticidal to at least European corn borer. Although both constructs produced transgenic events that were very active against at least ECB, generally transgenic plants with the zmUbi promoter driving expression of the TM33 coding sequence produced higher levels of eCry1Ba protein than transgenic plants comprising the MTL promoter. The eCry1Ba protein concentration ranged from 460-681  $\mu g/mg$  soluble protein for the 18319 construct and 509-2984  $\mu g/mg$  soluble protein for the 18320 construct.

TABLE 7

	Activ	ity of transs	genic maize expr	ressing eCry1Ba proteins
	Construct	Maize Event	ECB Activity	eCry1Ba Concentration (μg/mg soluble protein)
_	18319	9A	+	648
		23A	+	676
		38A	+	624
		<b>4</b> 0 <b>A</b>	+	460
		52A	+	681
		57A	+	618
	18320	28A	+	1839
		29B	+	2818
		34C	+	2984
		42A	+	1625
		46A	+	1010
		48B	+	509

Results shown in Table 8 indicate that transgenic sugarcane plants expressing the eCry1Ba-T2A:Y150K:L189S mutant protein are insecticidal to sugarcane borer.

TABLE 8

Activity of transgenic sugarcane	e expressing eCry1Ba and Vip3
Event	SCB
72581-1A	+
72581-2A 72581-3A	+
72581-4A	+

TABLE 8-continued

36 TABLE 9-continued

A	ctivity of transgenic sugarcan	e expressing eCry1Ba and Vip3		Co.	mparis
	Event	SCB	5	DOMARI	DEC
	72581-5A	_		DOMAIN	REC
	Control	-			V2 CB2
				II	СБ2
	F	1 10	10		V3 CB3

## Structure of Cry1Ba Protein

Example 10

Table 9 shows the relationship between the three domains 15 of Cry1Ab (SEQ ID NO:51) and Cry1Ba (SEQ ID NO:2) with their respective variable regions and conserved blocks. The amino acids comprised in each domain, conserved block and variable region is shown for both proteins.

TABLE 9

Cor	nparison of stru	cture of Cry1Ab	and Cry1Ba.
DOMAIN	REGION	Cry1Ab (FIG. 2)	Cry1Ba (SEQ ID NO: 2)
	V1	1-32	1-47
I	V1	33-152	48-171
	CB1	153-182	172-201

DOMAIN	REGION	Cry1Ab (FIG. 2)	Cry1Ba (SEQ ID NO: 2)
	V2	183-202	202-221
	CB2	203-254	222-270
II		255-269	271-288
	V3	270-452	289-480
	CB3	453-462	481-490
III		463-500	491-528
	V4	501-520	529-548
	CB4	521-531	549-559
	V5	532-596	560-624
	CB5	597-606	625-634
	V6	607-610	635-638
	Protoxin	611-1155	639-1228

It should be understood that the examples and embodiments described herein are for illustrative purposes only and 20 that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and the scope of the appended claims.

All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art that this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

SEQUENCE LISTING

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<223> OTHER INFORMATION: Native full-length cry1Ba coding sequence

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Tyr	Gln 130	Gln	Ser	Leu	Glu	Asp 135	Trp	Leu	Glu	Asn	Arg 140	Asp	Asp	Ala	Arg
Thr 145	Arg	Ser	Val	Leu	Lys 150	Thr	Gln	Tyr	Ile	Ala 155	Leu	Glu	Leu	Asp	Phe 160
Leu	Asn	Ala	Met	Pro 165	Leu	Phe	Ala	Ile	Arg 170	Asn	Gln	Glu	Val	Pro 175	Leu
Leu	Met	Val	Tyr 180	Ala	Gln	Ala	Ala	Asn 185	Leu	His	Leu	Leu	Leu 190	Leu	Arg
Asp	Ala	Ser 195	Leu	Phe	Gly	Ser	Glu 200	Phe	Gly	Leu	Thr	Ser 205	Gln	Glu	Ile
Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Asp	Tyr	Ser	Asp
Tyr 225	Сла	Val	Glu	Trp	Tyr 230	Asn	Thr	Gly	Leu	Asn 235	Ser	Leu	Arg	Gly	Thr 240
Asn	Ala	Ala	Ser	Trp 245	Val	Arg	Tyr	Asn	Gln 250	Phe	Arg	Arg	Asp	Leu 255	Thr
Leu	Gly	Val	Leu 260	Asp	Leu	Val	Ala	Leu 265	Phe	Pro	Ser	Tyr	Asp 270	Thr	Arg
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Asp	Ala 290	Ile	Gly	Ala	Thr	Gly 295	Val	Asn	Met	Ala	Ser 300	Met	Asn	Trp	Tyr
Asn 305	Asn	Asn	Ala	Pro	Ser 310	Phe	Ser	Ala	Ile	Glu 315	Ala	Ala	Ala	Ile	Arg 320
Ser	Pro	His	Leu	Leu 325	Asp	Phe	Leu	Glu	Gln 330	Leu	Thr	Ile	Phe	Ser 335	Ala
Ser	Ser	Arg	Trp 340	Ser	Asn	Thr	Arg	His 345	Met	Thr	Tyr	Trp	Arg 350	Gly	His
Thr	Ile	Gln 355	Ser	Arg	Pro	Ile	Gly 360	Gly	Gly	Leu	Asn	Thr 365	Ser	Thr	His

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Asn	Thr	Ala 115	Leu	Ala	Arg	Leu	Gln 120	Gly	Leu	Gly	Asp	Ser 125	Phe	Arg	Ala
Tyr	Gln 130	Gln	Ser	Leu	Glu	Asp 135	Trp	Leu	Glu	Asn	Arg 140	Asp	Asp	Ala	Arg
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Leu	Asn	Ala	Met	Pro 165	Leu	Phe	Ala	Ile	Arg 170	Asn	Gln	Glu	Val	Pro 175	Leu
Leu	Ser	Val	Tyr 180	Ala	Gln	Ala	Ala	Asn 185	Leu	His	Leu	Leu	Leu 190	Leu	Arg
Asp	Ala	Ser 195	Leu	Phe	Gly	Ser	Glu 200	Phe	Gly	Leu	Thr	Ser 205	Gln	Glu	Ile
Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Asp	Tyr	Ser	Asp
Tyr 225	Cys	Val	Glu	Trp	Tyr 230	Asn	Thr	Gly	Leu	Asn 235	Ser	Leu	Arg	Gly	Thr 240
Asn	Ala	Ala	Ser	Trp 245	Val	Arg	Tyr	Asn	Gln 250	Phe	Arg	Arg	Asp	Leu 255	Thr
Leu	Gly	Val	Leu 260	Asp	Leu	Val	Ala	Leu 265	Phe	Pro	Ser	Tyr	Asp 270	Thr	Arg
Thr	Tyr	Pro 275	Ile	Asn	Thr	Ser	Ala 280	Gln	Leu	Thr	Arg	Glu 285	Val	Tyr	Thr
Asp	Ala 290	Ile	Gly	Ala	Thr	Gly 295	Val	Asn	Met	Ala	Ser 300	Met	Asn	Trp	Tyr
Asn 305	Asn	Asn	Ala	Pro	Ser 310	Phe	Ser	Ala	Ile	Glu 315	Ala	Ala	Ala	Ile	Arg 320
Ser	Pro	His	Leu	Leu 325	Asp	Phe	Leu	Glu	Gln 330	Leu	Thr	Ile	Phe	Ser 335	Ala
Ser	Ser	Arg	Trp 340	Ser	Asn	Thr	Arg	His 345	Met	Thr	Tyr	Trp	Arg 350	Gly	His
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Thr	Asn	Pro	Gln 420	Asn	Ile	Ser	Asp	Arg 425	Gly	Thr	Ala	Asn	Tyr 430	Ser	Gln
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Tyr	Ser	Trp	Thr	His 485	Arg	Ser	Ala	Asp	Arg 490	Thr	Asn	Thr	Ile	Gly 495	Pro
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Gly 545	Pro	Leu	Thr	Gln	Arg 550	Tyr	Arg	Ile	Gly	Phe 555	Arg	Tyr	Ala	Ser	Thr 560
Val	Asp	Phe	Asp	Phe 565	Phe	Val	Ser	Arg	Gly 570	Gly	Thr	Thr	Val	Asn 575	Asn
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Asn	Phe	Val 595	Arg	Arg	Ala	Phe	Thr 600	Thr	Pro	Phe	Thr	Phe 605	Thr	Gln	Ile
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Val 625	Tyr	Ile	Asp	ГЛа	Ile 630	Glu	Ile	Ile	Pro	Val 635	Thr	Ala	Thr	Phe	Glu 640
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Tyr	Gln 130	Gln	Ser	Leu	Glu	Asp 135	Trp	Leu	Glu	Asn	Arg 140	Asp	Asp	Ala	Arg
Thr 145	Arg	Ser	Val	Leu	Lys 150	Thr	Gln	Tyr	Ile	Ala 155	Leu	Glu	Leu	Asp	Phe 160
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Leu	Met	Val	Tyr 180	Ala	Gln	Ala	Ala	Asn 185	Leu	His	Leu	Ser	Leu 190	Leu	Arg
Asp	Ala	Ser 195	Leu	Phe	Gly	Ser	Glu 200	Phe	Gly	Leu	Thr	Ser 205	Gln	Glu	Ile
Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Asp	Tyr	Ser	Asp
Tyr	Cys	Val	Glu	Trp	Tyr	Asn	Thr	Gly	Leu	Asn	Ser	Leu	Arg	Gly	Thr

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Thr	Tyr	Pro 275	Ile	Asn	Thr	Ser	Ala 280	Gln	Leu	Thr	Arg	Glu 285	Val	Tyr	Thr
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Tyr	Ser	Trp	Thr	His 485	Arg	Ser	Ala	Asp	Arg 490	Thr	Asn	Thr	Ile	Gly 495	Pro
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Gly 545	Pro	Leu	Thr	Gln	Arg 550	Tyr	Arg	Ile	Gly	Phe 555	Arg	Tyr	Ala	Ser	Thr 560
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Asn	Phe	Val 595	Arg	Arg	Ala	Phe	Thr 600	Thr	Pro	Phe	Thr	Phe 605	Thr	Gln	Ile
Gln	Asp 610	Ile	Ile	Arg	Thr	Ser 615	Ile	Gln	Gly	Leu	Ser 620	Gly	Asn	Gly	Glu
Val 625	Tyr	Ile	Asp	Lys	Ile 630	Glu	Ile	Ile	Pro	Val 635	Thr	Ala	Thr	Phe	Glu 640
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Ser	Thr 50	Val	Gln	Thr	Gly	Ile 55	Asn	Ile	Ala	Gly	Arg 60	Ile	Leu	Gly	Val
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Val	Gly	Glu	Leu	Trp 85	Pro	Arg	Gly	Arg	Asp 90	Gln	Trp	Glu	Ile	Phe 95	Leu
Glu	His	Val	Glu 100	Gln	Leu	Ile	Asn	Gln 105	Gln	Ile	Thr	Glu	Asn 110	Ala	Arg
Asn	Thr	Ala 115	Leu	Ala	Arg	Leu	Gln 120	Gly	Leu	Gly	Asp	Ser 125	Phe	Arg	Ala
Tyr	Gln 130	Gln	Ser	Leu	Glu	Asp 135	Trp	Leu	Glu	Asn	Arg 140	Asp	Asp	Ala	Arg
Thr 145	Arg	Ser	Val	Leu	Lys 150	Thr	Gln	Tyr	Ile	Ala 155	Leu	Glu	Leu	Asp	Phe 160
Leu	Asn	Ala	Met	Pro 165	Leu	Phe	Ala	Ile	Arg 170	Asn	Gln	Glu	Val	Pro 175	Leu
Leu	Met	Val	Tyr 180	Ala	Gln	Ala	Ala	Asn 185	Leu	His	Leu	Leu	Leu 190	Leu	Arg
Asp	Ala	Ser 195	Leu	Phe	Gly	Lys	Glu 200	Phe	Gly	Leu	Thr	Ser 205	Gln	Glu	Ile
Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Asp	Tyr	Ser	Asp
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Ser	Pro	His	Leu	Leu 325	Asp	Phe	Leu	Glu	Gln 330	Leu	Thr	Ile	Phe	Ser 335	Ala
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Asn	Ala	Ala	Ser	Trp 245	Val	Arg	Tyr	Asn	Gln 250	Phe	Arg	Arg	Asp	Leu 255	Thr
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Asn Arg Ile Thr Gln Ile Pro Met Val Lys Ala Ser Glu Leu Pro Gln

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Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Asp	Tyr	Ser	Asp
Tyr 225	Cys	Val	Glu	Trp	Tyr 230	Asn	Thr	Gly	Leu	Asn 235	Ser	Leu	Arg	Gly	Thr 240
Asn	Ala	Ala	Ser	Trp 245	Val	Arg	Tyr	Asn	Gln 250	Phe	Arg	Arg	Asp	Leu 255	Thr
Leu	Gly	Val	Leu 260	Asp	Leu	Val	Ala	Leu 265	Phe	Pro	Ser	Tyr	Asp 270	Thr	Arg
Thr	Tyr	Pro 275	Ile	Asn	Thr	Ser	Ala 280	Gln	Leu	Thr	Arg	Glu 285	Val	Tyr	Thr
Asp	Ala 290	Ile	Gly	Ala	Thr	Gly 295	Val	Asn	Met	Ala	Ser 300	Met	Asn	Trp	Tyr
Asn 305	Asn	Asn	Ala	Pro	Ser 310	Phe	Ser	Ala	Ile	Glu 315	Ala	Ala	Ala	Ile	Arg 320
Ser	Pro	His	Leu	Leu 325	Asp	Phe	Leu	Glu	Gln 330	Leu	Thr	Ile	Phe	Ser 335	Ala
Ser	Ser	Arg	Trp 340	Ser	Asn	Thr	Arg	His 345	Met	Thr	Tyr	Trp	Arg 350	Gly	His
Thr	Ile	Gln 355	Ser	Arg	Pro	Ile	Gly 360	Gly	Gly	Leu	Asn	Thr 365	Ser	Thr	His
Gly	Ala 370	Thr	Asn	Thr	Ser	Ile 375	Asn	Pro	Val	Thr	Leu 380	Arg	Phe	Ala	Ser
Arg 385	Asp	Val	Tyr	Arg	Thr 390	Glu	Ser	Tyr	Ala	Gly 395	Val	Leu	Leu	Trp	Gly 400
Ile	Tyr	Leu	Glu	Pro 405	Ile	His	Gly	Val	Pro 410	Thr	Val	Arg	Phe	Asn 415	Phe
Thr	Asn	Pro	Gln 420	Asn	Ile	Ser	Asp	Arg 425	Gly	Thr	Ala	Asn	Tyr 430	Ser	Gln
Pro	Tyr	Glu 435	Ser	Pro	Gly	Leu	Gln 440	Leu	Lys	Asp	Ser	Glu 445	Thr	Glu	Leu
Pro	Pro 450	Glu	Thr	Thr	Glu	Arg 455	Pro	Asn	Tyr	Glu	Ser 460	Tyr	Ser	His	Arg
Leu 465	Ser	His	Ile	Gly	Ile 470	Ile	Leu	Gln	Ser	Arg 475	Val	Asn	Val	Pro	Val 480
Tyr	Ser	Trp	Thr	His 485	Arg	Ser	Ala	Asp	Arg 490	Thr	Asn	Thr	Ile	Gly 495	Pro
Asn	Arg		Thr 500		Ile	Pro		Val 505		Ala	Ser	Glu	Leu 510	Pro	Gln
Gly	Thr	Thr 515	Val	Val	Arg	Gly	Pro 520	Gly	Phe	Thr	Gly	Gly 525	Asp	Ile	Leu
Arg	Arg 530	Thr	Asn	Thr	Gly	Gly 535	Phe	Gly	Pro	Ile	Arg 540	Val	Thr	Val	Asn
Gly 545	Pro	Leu	Thr	Gln	Arg 550	Tyr	Arg	Ile	Gly	Phe 555	Arg	Tyr	Ala	Ser	Thr 560
Val	Asp	Phe	Asp	Phe 565	Phe	Val	Ser	Arg	Gly 570	Gly	Thr	Thr	Val	Asn 575	Asn
Phe	Arg	Phe	Leu 580	Arg	Thr	Met	Asn	Ser 585	Gly	Asp	Glu	Leu	Lys 590	Tyr	Gly
Asn	Phe	Val 595	Arg	Arg	Ala	Phe	Thr 600	Thr	Pro	Phe	Thr	Phe 605	Thr	Gln	Ile
Gln	Asp 610	Ile	Ile	Arg	Thr	Ser 615	Ile	Gln	Gly	Leu	Ser 620	Gly	Asn	Gly	Glu
Val	Tyr	Ile	Asp	rys	Ile	Glu	Ile	Ile	Pro	Val	Thr	Ala	Thr	Phe	Glu

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<213 > ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Maize-optimized sequence encoding eCry1Ba-T2A,Y150K,L189S mutant

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<223> OTHER INFORMATION: s= g or c
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atatgtttaa acatgacttc aaataggaaa aatgagaatg aa
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<220> FEATURE:
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<223> OTHER INFORMATION: YG163 forward primer
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<212> TYPE: DNA
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<220> FEATURE:
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<220> FEATURE:
<223> OTHER INFORMATION: YG175 primer
<400> SEQUENCE: 29
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<212> TYPE: DNA
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<223 > OTHER INFORMATION: YG176 primer
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<212> TYPE: DNA
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<220> FEATURE:
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<223> OTHER INFORMATION: YG193 reverse primer
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<212> TYPE: PRT
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<222> LOCATION: (1)..(626)
<223 > OTHER INFORMATION: Truncated Cry1Ba-T7
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Gly Ile Asn Ile Ala Gly Arg Ile Leu Gly Val Leu Gly Val Pro Phe
                            40
Ala Gly Gln Leu Ala Ser Phe Tyr Ser Phe Leu Val Gly Glu Leu Trp
             55
Pro Arg Gly Arg Asp Gln Trp Glu Ile Phe Leu Glu His Val Glu Gln
                    70
Leu Ile Asn Gln Gln Ile Thr Glu Asn Ala Arg Asn Thr Ala Leu Ala
                                    90
Arg Leu Gln Gly Leu Gly Asp Ser Phe Arg Ala Tyr Gln Gln Ser Leu
          100
                            105
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Tyr	Thr 130	Gln	Tyr	Ile	Ala	Leu 135	Glu	Leu	Asp	Phe	Leu 140	Asn	Ala	Met	Pro
Leu 145	Phe	Ala	Ile	Arg	Asn 150	Gln	Glu	Val	Pro	Leu 155	Leu	Met	Val	Tyr	Ala 160
Gln	Ala	Ala	Asn	Leu 165	His	Leu	Leu	Leu	Leu 170	Arg	Asp	Ala	Ser	Leu 175	Phe
Gly	Ser	Glu	Phe 180	Gly	Leu	Thr	Ser	Gln 185	Glu	Ile	Gln	Arg	Tyr 190	Tyr	Glu
Arg	Gln	Val 195	Glu	Arg	Thr	Arg	Asp 200	Tyr	Ser	Asp	Tyr	Сув 205	Val	Glu	Trp
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Val 225	Arg	Tyr	Asn	Gln	Phe 230	Arg	Arg	Asp	Leu	Thr 235	Leu	Gly	Val	Leu	Asp 240
Leu	Val	Ala	Leu	Phe 245	Pro	Ser	Tyr	Asp	Thr 250	Arg	Thr	Tyr	Pro	Ile 255	Asn
Thr	Ser	Ala	Gln 260	Leu	Thr	Arg	Glu	Val 265	Tyr	Thr	Asp	Ala	Ile 270	Gly	Ala
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Thr	Glu 370	Ser	Tyr	Ala	Gly	Val 375	Leu	Leu	Trp	Gly	Ile 380	Tyr	Leu	Glu	Pro
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Ile	Ser	Asp	Arg	Gly 405	Thr	Ala	Asn	Tyr	Ser 410	Gln	Pro	Tyr	Glu	Ser 415	Pro
Gly	Leu	Gln	Leu 420	ГÀа	Asp	Ser	Glu	Thr 425	Glu	Leu	Pro	Pro	Glu 430	Thr	Thr
Glu	Arg	Pro 435	Asn	Tyr	Glu	Ser	Tyr 440	Ser	His	Arg	Leu	Ser 445	His	Ile	Gly
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Arg 465	Ser	Ala	Asp	Arg	Thr 470	Asn	Thr	Ile	Gly	Pro 475	Asn	Arg	Ile	Thr	Gln 480
Ile	Pro	Met	Val	Lys 485	Ala	Ser	Glu	Leu	Pro 490	Gln	Gly	Thr	Thr	Val 495	Val
Arg	Gly	Pro	Gly 500	Phe	Thr	Gly	Gly	Asp 505	Ile	Leu	Arg	Arg	Thr 510	Asn	Thr
Gly	Gly	Phe 515	Gly	Pro	Ile	Arg	Val 520	Thr	Val	Asn	Gly	Pro 525	Leu	Thr	Gln

												COII	C III.	aca	
Arg	Tyr 530	Arg	Ile	Gly	Phe	Arg 535	Tyr	Ala	Ser	Thr	Val 540	Asp	Phe	Asp	Phe
Phe 545	Val	Ser	Arg	Gly	Gly 550	Thr	Thr	Val	Asn	Asn 555	Phe	Arg	Phe	Leu	Arg 560
Thr	Met	Asn	Ser	Gly 565	Asp	Glu	Leu	Lys	Tyr 570	Gly	Asn	Phe	Val	Arg 575	Arg
Ala	Phe	Thr	Thr 580	Pro	Phe	Thr	Phe	Thr 585	Gln	Ile	Gln	Asp	Ile 590	Ile	Arg
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Glu 625	Arg														
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Ser	Leu	Сув 35	Ile	Ala	Glu	Gly	Asn 40	Asn	Ile	Asp	Pro	Phe 45	Val	Ser	Ala
Ser	Thr 50	Val	Gln	Thr	Gly	Ile 55	Asn	Ile	Ala	Gly	Arg 60	Ile	Leu	Gly	Val
Leu 65	Gly	Val	Pro	Phe	Ala 70	Gly	Gln	Leu	Ala	Ser 75	Phe	Tyr	Ser	Phe	Leu 80
Val	Gly	Glu	Leu	Trp 85	Pro	Arg	Gly	Arg	Asp 90	Gln	Trp	Glu	Ile	Phe 95	Leu
Glu	His	Val	Glu 100	Gln	Leu	Ile	Asn	Gln 105	Gln	Ile	Thr	Glu	Asn 110	Ala	Arg
Asn	Thr	Ala 115	Leu	Ala	Arg	Leu	Gln 120	Gly	Leu	Gly	Asp	Ser 125	Phe	Arg	Ala
Tyr	Gln 130	Gln	Ser	Leu	Glu	Asp 135	Trp	Leu	Glu	Asn	Arg 140	Asp	Asp	Ala	Arg
Thr 145	Arg	Ser	Val	Leu	Lys 150	Thr	Gln	Tyr	Ile	Ala 155	Leu	Glu	Leu	Asp	Phe 160
Leu	Asn	Ala	Met	Pro 165	Leu	Phe	Ala	Ile	Arg 170	Asn	Gln	Glu	Val	Pro 175	Leu
Leu	Pro	Val	Tyr 180	Ala	Gln	Ala	Ala	Asn 185	Leu	His	Leu	Leu	Leu 190	Leu	Arg
Asp	Ala	Ser 195	Leu	Phe	Gly	Ser	Glu 200	Phe	Gly	Leu	Thr	Ser 205	Gln	Glu	Ile
Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Asp	Tyr	Ser	Asp
Tyr 225	Сув	Val	Glu	Trp	Tyr 230	Asn	Thr	Gly	Leu	Asn 235	Ser	Leu	Arg	Gly	Thr 240
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Asp	Ala 290	Ile	Gly	Ala	Thr	Gly 295	Val	Asn	Met	Ala	Ser 300	Met	Asn	Trp	Tyr
Asn 305	Asn	Asn	Ala	Pro	Ser 310	Phe	Ser	Ala	Ile	Glu 315	Ala	Ala	Ala	Ile	Arg 320
Ser	Pro	His	Leu	Leu 325	Asp	Phe	Leu	Glu	Gln 330	Leu	Thr	Ile	Phe	Ser 335	Ala
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Thr	Ile	Gln 355	Ser	Arg	Pro	Ile	Gly 360	Gly	Gly	Leu	Asn	Thr 365	Ser	Thr	His
Gly	Ala 370	Thr	Asn	Thr	Ser	Ile 375	Asn	Pro	Val	Thr	Leu 380	Arg	Phe	Ala	Ser
Arg 385	Asp	Val	Tyr	Arg	Thr 390	Glu	Ser	Tyr	Ala	Gly 395	Val	Leu	Leu	Trp	Gly 400
Ile	Tyr	Leu	Glu	Pro 405	Ile	His	Gly	Val	Pro 410	Thr	Val	Arg	Phe	Asn 415	Phe
Thr	Asn	Pro	Gln 420	Asn	Ile	Ser	Asp	Arg 425	Gly	Thr	Ala	Asn	Tyr 430	Ser	Gln
Pro	Tyr	Glu 435	Ser	Pro	Gly	Leu	Gln 440	Leu	Lys	Asp	Ser	Glu 445	Thr	Glu	Leu
Pro	Pro 450	Glu	Thr	Thr	Glu	Arg 455	Pro	Asn	Tyr	Glu	Ser 460	Tyr	Ser	His	Arg
Leu 465	Ser	His	Ile	Gly	Ile 470	Ile	Leu	Gln	Ser	Arg 475	Val	Asn	Val	Pro	Val 480
Tyr	Ser	Trp	Thr	His 485	Arg	Ser	Ala	Asp	Arg 490	Thr	Asn	Thr	Ile	Gly 495	Pro
Asn	Arg	Ile	Thr 500	Gln	Ile	Pro	Met	Val 505	Lys	Ala	Ser	Glu	Leu 510	Pro	Gln
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Gly 545	Pro	Leu	Thr	Gln	Arg 550	Tyr	Arg	Ile	Gly	Phe 555	Arg	Tyr	Ala	Ser	Thr 560
Val	Asp	Phe	Asp	Phe 565	Phe	Val	Ser	Arg	Gly 570	Gly	Thr	Thr	Val	Asn 575	Asn
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Gln	Asp 610	Ile	Ile	Arg	Thr	Ser 615	Ile	Gln	Gly	Leu	Ser 620	Gly	Asn	Gly	Glu
Val 625	Tyr	Ile	Asp	Lys	Ile 630	Glu	Ile	Ile	Pro	Val 635	Thr	Ala	Thr	Phe	Glu 640
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Ser	Leu	Сув 35	Ile	Ala	Glu	Gly	Asn 40	Asn	Ile	Asp	Pro	Phe 45	Val	Ser	Ala
Ser	Thr 50	Val	Gln	Thr	Gly	Ile 55	Asn	Ile	Ala	Gly	Arg 60	Ile	Leu	Gly	Val
Leu 65	Gly	Val	Pro	Phe	Ala 70	Gly	Gln	Leu	Ala	Ser 75	Phe	Tyr	Ser	Phe	Leu 80
Val	Gly	Glu	Leu	Trp 85	Pro	Arg	Gly	Arg	Asp 90	Gln	Trp	Glu	Ile	Phe 95	Leu
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Tyr	Gln 130	Gln	Ser	Leu	Glu	Asp 135	Trp	Leu	Glu	Asn	Arg 140	Asp	Asp	Ala	Arg
Thr 145	Arg	Ser	Val	Leu	Lys 150	Thr	Gln	Tyr	Ile	Ala 155	Leu	Glu	Leu	Asp	Phe 160
Leu	Asn	Ala	Met	Pro 165	Leu	Phe	Ala	Ile	Ser 170	Asn	Gln	Glu	Val	Pro 175	Leu
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Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Asp	Tyr	Ser	Asp
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Asn	Ala	Ala	Ser	Trp 245	Val	Arg	Tyr	Asn	Gln 250	Phe	Arg	Arg	Asp	Leu 255	Thr
Leu	Gly	Val	Leu 260	Asp	Leu	Val	Ala	Leu 265	Phe	Pro	Ser	Tyr	Asp 270	Thr	Arg
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Ser	Pro	His	Leu	Leu 325	Asp	Phe	Leu	Glu	Gln 330	Leu	Thr	Ile	Phe	Ser 335	Ala
Ser	Ser	Arg	Trp 340	Ser	Asn	Thr	Arg	His 345	Met	Thr	Tyr	Trp	Arg 350	Gly	His
Thr	Ile	Gln 355	Ser	Arg	Pro	Ile	Gly 360	Gly	Gly	Leu	Asn	Thr 365	Ser	Thr	His
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Tyr	Ser	Trp	Thr	His 485	Arg	Ser	Ala	Asp	Arg 490	Thr	Asn	Thr	Ile	Gly 495	Pro
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Gly 545	Pro	Leu	Thr	Gln	Arg 550	Tyr	Arg	Ile	Gly	Phe 555	Arg	Tyr	Ala	Ser	Thr 560
Val	Asp	Phe	Asp	Phe 565	Phe	Val	Ser	Arg	Gly 570	Gly	Thr	Thr	Val	Asn 575	Asn
Phe	Arg	Phe	Leu 580	Arg	Thr	Met	Asn	Ser 585	Gly	Asp	Glu	Leu	590 Lys	Tyr	Gly
Asn	Phe	Val 595	Arg	Arg	Ala	Phe	Thr 600	Thr	Pro	Phe	Thr	Phe 605	Thr	Gln	Ile
Gln	Asp 610	Ile	Ile	Arg	Thr	Ser 615	Ile	Gln	Gly	Leu	Ser 620	Gly	Asn	Gly	Glu
Val 625	Tyr	Ile	Asp	Lys	Ile 630	Glu	Ile	Ile	Pro	Val 635	Thr	Ala	Thr	Phe	Glu 640
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Ser	Leu	Cys	Ile	Ala	Glu	Gly	Asn 40	Asn	Ile	Asp	Pro	Phe 45	Val	Ser	Ala
Ser	Thr 50	Val	Gln	Thr	Gly	Ile 55	Asn	Ile	Ala	Gly	Arg 60	Ile	Leu	Gly	Val
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Val	Gly	Glu	Leu	Trp 85	Pro	Arg	Gly	Arg	Asp	Gln	Trp	Glu	Ile	Phe 95	Leu
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Leu	Asn	Ala	Met	Pro 165	Leu	Phe	Ala	Ile	Arg 170	Asn	Gln	Glu	Val	Pro 175	Leu
Leu	Met	Val	Tyr 180	Ala	Gln	Ala	Ala	Asn 185	Leu	His	Leu	Leu	Leu 190	Leu	Arg
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Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Asp	Tyr	Ser	Asp
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Asn	Ala	Ala	Ser	Trp 245	Val	Arg	Tyr	Asn	Gln 250	Phe	Arg	Arg	Asp	Leu 255	Thr
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Ser	Pro	His	Leu	Leu 325	Asp	Phe	Leu	Glu	Gln 330	Leu	Thr	Ile	Phe	Ser 335	Ala
Ser	Ser	Arg	Trp 340	Ser	Asn	Thr	Arg	His 345	Met	Thr	Tyr	Trp	Arg 350	Gly	His
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Leu 465	Ser	His	Ile	Gly	Ile 470	Ile	Leu	Gln	Ser	Arg 475	Val	Asn	Val	Pro	Val 480
Tyr	Ser	Trp	Thr	His 485	Arg	Ser	Ala	Asp	Arg 490	Thr	Asn	Thr	Ile	Gly 495	Pro
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Val 625	Tyr	Ile	Asp	ГÀа	Ile 630	Glu	Ile	Ile	Pro	Val 635	Thr	Ala	Thr	Phe	Glu 640
Ala	Glu	Tyr	Asp	Leu 645	Glu	Arg	Ala	Gln	Glu 650	Ala	Val	Asn	Ala	Leu 655	Phe
Thr	Asn	Thr	Asn 660	Pro	Arg	Arg	Leu	Lys 665	Thr	Asp	Val	Thr	Asp 670	Tyr	His
Ile	Asp	Gln 675	Val	Ser	Asn	Leu	Val 680	Ala	Cys	Leu	Ser	Asp 685	Glu	Phe	Cys
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Asn	Lys	Gln	Pro	Asp 725	Phe	Ile	Ser	Thr	Asn 730	Glu	Gln	Ser	Asn	Phe 735	Thr
Ser	Ile	His	Glu 740	Gln	Ser	Glu	His	Gly 745	Trp	Trp	Gly	Ser	Glu 750	Asn	Ile
Thr	Ile	Gln 755	Glu	Gly	Asn	Asp	Val 760	Phe	Lys	Glu	Asn	Tyr 765	Val	Thr	Leu
Pro	Gly 770	Thr	Phe	Asn	Glu	Суs 775	Tyr	Pro	Thr	Tyr	Leu 780	Tyr	Gln	ГÀЗ	Ile
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Lys	His	Glu	Thr 820	Leu	Asp	Val	Pro	Gly 825	Thr	Glu	Ser	Leu	Trp 830	Pro	Leu
Ser	Val	Glu 835	Ser	Pro	Ile	Gly	Arg 840	CÀa	Gly	Glu	Pro	Asn 845	Arg	CÀa	Ala
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Glu 865	Lys	Cys	Ala	His	His 870	Ser	His	His	Phe	Ser 875	Leu	Asp	Ile	Asp	Val 880
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Asn	Phe	Val 595	Arg	Arg	Ala	Phe	Thr 600	Thr	Pro	Phe	Thr	Phe 605	Thr	Gln	Ile
Gln	Asp 610	Ile	Ile	Arg	Thr	Ser 615	Ile	Gln	Gly	Leu	Ser 620	Gly	Asn	Gly	Glu
Val 625	Tyr	Ile	Asp	ГЛа	Ile 630	Glu	Ile	Ile	Pro	Val 635	Thr	Ala	Thr	Phe	Glu 640
Ala	Glu	Tyr	Asp	Leu 645	Glu	Arg	Ala	Gln	Glu 650	Ala	Val	Asn	Ala	Leu 655	Phe
Thr	Asn	Thr	Asn 660	Pro	Arg	Arg	Leu	Lys	Thr	Asp	Val	Thr	Asp 670	Tyr	His
Ile	Asp	Gln 675	Val	Ser	Asn	Leu	Val 680	Ala	Cys	Leu	Ser	Asp 685	Glu	Phe	CÀa
Leu	Asp 690	Glu	Lys	Arg	Glu	Leu 695	Leu	Glu	Lys	Val	Lys 700	Tyr	Ala	ГЛа	Arg
Leu 705	Ser	Asp	Glu	Arg	Asn 710	Leu	Leu	Gln	Asp	Pro 715	Asn	Phe	Thr	Ser	Ile 720
Asn	Lys	Gln	Pro	Asp 725	Phe	Ile	Ser	Thr	Asn 730	Glu	Gln	Ser	Asn	Phe 735	Thr
Ser	Ile	His	Glu 740	Gln	Ser	Glu	His	Gly 745	Trp	Trp	Gly	Ser	Glu 750	Asn	Ile
Thr	Ile	Gln 755	Glu	Gly	Asn	Asp	Val 760	Phe	Lys	Glu	Asn	Tyr 765	Val	Thr	Leu
Pro	Gly 770	Thr	Phe	Asn	Glu	Сув 775	Tyr	Pro	Thr	Tyr	Leu 780	Tyr	Gln	Lys	Ile
Gly 785	Glu	Ser	Glu	Leu	Lys 790	Ala	Tyr	Thr	Arg	Tyr 795	Gln	Leu	Arg	Gly	Tyr 800
Ile	Glu	Asp	Ser	Gln 805	Asp	Leu	Glu	Ile	Tyr 810	Leu	Ile	Arg	Tyr	Asn 815	Ala
Lys	His	Glu	Thr 820	Leu	Asp	Val	Pro	Gly 825	Thr	Glu	Ser	Leu	Trp 830	Pro	Leu
Ser	Val	Glu 835	Ser	Pro	Ile	Gly	Arg 840	Cya	Gly	Glu	Pro	Asn 845	Arg	Càa	Ala
Pro	His 850	Phe	Glu	Trp	Asn	Pro 855	Asp	Leu	Asp	Cys	Ser 860	Cys	Arg	Asp	Gly
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Asn	Thr	Ala 115	Leu	Ala	Arg	Leu	Gln 120	Gly	Leu	Gly	Asp	Ser 125	Phe	Arg	Ala
Tyr	Gln 130	Gln	Ser	Leu	Glu	Asp 135	Trp	Leu	Glu	Asn	Arg 140	Asp	Asp	Ala	Arg
Thr 145	Arg	Ser	Val	Leu	Tyr 150	Thr	Gln	Tyr	Ile	Ala 155	Leu	Glu	Leu	Asp	Phe 160
Leu	Asn	Ala	Met	Pro 165	Leu	Phe	Ala	Ile	Arg 170	Asn	Gln	Glu	Val	Pro 175	Leu
Leu	Met	Val	Tyr 180	Ala	Gln	Ala	Ala	Asn 185	Leu	His	Leu	Leu	Leu 190	Leu	Arg
Asp	Ala	Ser 195	Leu	Phe	Gly	Ser	Glu 200	Phe	Gly	Leu	Thr	Ser 205	Gln	Glu	Ile
Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Aap	Tyr	Ser	Asp
Tyr 225	Cys	Val	Glu	Trp	Tyr 230	Asn	Thr	Gly	Leu	Asn 235	Ser	Leu	Arg	Gly	Thr 240
Asn	Ala	Ala	Ser	Trp 245	Val	Arg	Tyr	Asn	Gln 250	Phe	Arg	Arg	Asp	Leu 255	Thr
Leu	Gly	Val	Leu 260	Asp	Leu	Val	Ala	Leu 265	Phe	Pro	Ser	Tyr	Asp 270	Thr	Arg
Thr	Tyr	Pro 275	Ile	Asn	Thr	Ser	Ala 280	Gln	Leu	Thr	Arg	Glu 285	Val	Tyr	Thr
Asp	Ala 290	Ile	Gly	Ala	Thr	Gly 295	Val	Asn	Met	Ala	Ser 300	Met	Asn	Trp	Tyr
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Ser	Pro	His	Leu	Leu 325	Asp	Phe	Leu	Glu	Gln 330	Leu	Thr	Ile	Phe	Ser 335	Ala
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Pro	Pro 450	Glu	Thr	Thr	Glu	Arg 455	Pro	Asn	Tyr	Glu	Ser 460	Tyr	Ser	His	Arg
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Gly	Thr	Thr 515	Val	Val	Arg	Gly	Pro 520	Gly	Phe	Thr	Gly	Gly 525	Asp	Ile	Leu
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Val 625	Tyr	Ile	Asp	Lys	Ile 630	Glu	Ile	Ile	Pro	Val 635	Thr	Ala	Thr	Phe	Glu 640
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Thr	Asn	Thr	Asn 660	Pro	Arg	Arg	Leu	665 665	Thr	Asp	Val	Thr	Asp 670	Tyr	His
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Thr	Ile	Gln 755	Glu	Gly	Asn	Asp	Val 760	Phe	Lys	Glu	Asn	Tyr 765	Val	Thr	Leu
Pro	Gly 770	Thr	Phe	Asn	Glu	Сув 775	Tyr	Pro	Thr	Tyr	Leu 780	Tyr	Gln	Lys	Ile
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Ser	Val	Glu 835	Ser	Pro	Ile	Gly	Arg 840	Сув	Gly	Glu	Pro	Asn 845	Arg	Сув	Ala
Pro	His 850	Phe	Glu	Trp	Asn	Pro 855	Asp	Leu	Asp	СЛа	Ser 860	СЛа	Arg	Asp	Gly
Glu 865	Lys	Cys	Ala	His	His 870	Ser	His	His	Phe	Ser 875	Leu	Asp	Ile	Asp	Val 880
Gly	Cys	Thr	Asp	Leu 885	His	Glu	Asn	Leu	Gly 890	Val	Trp	Val	Val	Phe 895	ГХа
Ile	Lys	Thr	Gln	Glu	Gly	His	Ala	Arg	Leu	Gly	Asn	Leu	Glu	Phe	Ile

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			900					905					910	)	
Glu	Glu	Lys 915	Pro	Leu	Leu	Gly	Glu 920	Ala	Leu	Ser	Arg	Val 925	_	a Arg	g Ala
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Val	Lys 1025		n Gly	y Asp	Phe	Asr 103		∍n G	ly Le	eu Th		ys 035	Trp	Asn	Val
Lys	Gly 1040		g Val	l Asp	Val	. Glr 104		ln Se	er H	is Hi		rg 050	Ser	Asp	Leu
Val	Ile 1055		Gl:	ı Trg	Glu	106		Lu Va	al Se	er Gl		la 065	Val	Arg	Val
CAa	Pro 1070		ү Су:	∃ Gl∑	7 Tyr	107		∍u Ai	rg Va	al Th		la 080	Tyr	Lys	Glu
Gly	Tyr 1085		/ Gl	ı Gly	/ Cys	Val		ır I	le H:	is Gl		le 095	Glu	Asn	Asn
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Pro	Thr 1115		Th:	r Gly	/ Thr	Cys 112		an As	ap Ty	yr Th		la 125	His	Gln	Gly
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Glu	Asp 1145		а Ту:	r Glu	ı Val	. Asp		ır Tl	nr A	la Se		al 155	Asn	Tyr	Lys
Pro	Thr 1160		r Gli	ı Glu	ı Glu	116		/r Tl	nr As	sp Va		rg 170	Arg	Asp	Asn
	Cys 1179		и Ту:	r As <u>r</u>	Arç	Gl <sub>y</sub> 118	7 T3 30	/r Va	al As	sn Ty				Val	Pro
Ala	Gly 1190	_	r Vai	l Thi	. Lys	Glu 119		eu G	lu Ty	yr Ph		ro 200	Glu	Thr	Asp
Thr	Val 1205		, Ile	e Glu	ı Ile	Gl <sub>y</sub> 121		iu Tl	nr G	lu Gl		ys 215	Phe	Ile	Val
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Leu	Asn	Ala	Met	Pro 165	Leu	Phe	Ala	Ile	Arg 170	Asn	Gln	Glu	Val	Pro 175	Leu
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Asp	Ala	Ser 195	Leu	Phe	Gly	Ser	Glu 200	Phe	Gly	Leu	Thr	Ser 205	Gln	Glu	Ile
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Ile	Tyr	Leu	Glu	Pro 405	Ile	His	Gly	Val	Pro 410	Thr	Val	Arg	Phe	Asn 415	Phe
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Pro	Tyr	Glu 435	Ser	Pro	Gly	Leu	Gln 440	Leu	ГЛа	Asp	Ser	Glu 445	Thr	Glu	Leu
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Thr	Ile	Gln 755	Glu	Gly	Asn	Asp	Val 760	Phe	Lys	Glu	Asn	Tyr 765	Val	Thr	Leu
Pro	Gly 770	Thr	Phe	Asn	Glu	Сув 775	Tyr	Pro	Thr	Tyr	Leu 780	Tyr	Gln	Lys	Ile
Gly 785	Glu	Ser	Glu	Leu	Lys 790	Ala	Tyr	Thr	Arg	Tyr 795	Gln	Leu	Arg	Gly	Tyr 800
Ile	Glu	Asp	Ser	Gln 805	Asp	Leu	Glu	Ile	Tyr 810	Leu	Ile	Arg	Tyr	Asn 815	Ala
ГÀв	His	Glu	Thr 820	Leu	Asp	Val	Pro	Gly 825	Thr	Glu	Ser	Leu	Trp 830	Pro	Leu
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Pro	His 850	Phe	Glu	Trp	Asn	Pro 855	Asp	Leu	Asp	Сув	Ser 860	Сув	Arg	Asp	Gly
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Ser	Leu	Сув 35	Ile	Ala	Glu	Gly	Asn 40	Asn	Ile	Asp	Pro	Phe 45	Val	Ser	Ala
Ser	Thr 50	Val	Gln	Thr	Gly	Ile 55	Asn	Ile	Ala	Gly	Arg 60	Ile	Leu	Gly	Val
Leu 65	Gly	Val	Pro	Phe	Ala 70	Gly	Gln	Leu	Ala	Ser 75	Phe	Tyr	Ser	Phe	Leu 80
Val	Gly	Glu	Leu	Trp 85	Pro	Arg	Gly	Arg	Asp 90	Gln	Trp	Glu	Ile	Phe 95	Leu
Glu	His	Val	Glu 100	Gln	Leu	Ile	Asn	Gln 105	Gln	Ile	Thr	Glu	Asn 110	Ala	Arg
Asn	Thr	Ala 115	Leu	Ala	Arg	Leu	Gln 120	Gly	Leu	Gly	Asp	Ser 125	Phe	Arg	Ala
Tyr	Gln 130	Gln	Ser	Leu	Glu	Asp 135	Trp	Leu	Glu	Asn	Arg 140	Asp	Asp	Ala	Arg
Thr 145	Arg	Ser	Val	Leu	Tyr 150	Thr	Gln	Tyr	Ile	Ala 155	Leu	Glu	Leu	Asp	Phe 160
Leu	Asn	Ala	Met	Pro 165	Leu	Phe	Ala	Ile	Arg 170	Asn	Gln	Glu	Val	Pro 175	Leu
Leu	Met	Val	Tyr 180	Ala	Gln	Ala	Ala	Asn 185	Leu	His	Leu	Leu	Leu 190	Leu	Arg
Asp	Ala	Ser 195	Leu	Phe	Gly	Ser	Glu 200	Phe	Gly	Leu	Thr	Ser 205	Gln	Glu	Ile
Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Asp	Tyr	Ser	Asp
Tyr 225	Cha	Val	Glu	Trp	Tyr 230	Asn	Thr	Gly	Leu	Asn 235	Ser	Leu	Arg	Gly	Thr 240
Asn	Ala	Ala	Ser	Trp 245	Val	Arg	Tyr	Asn	Gln 250	Phe	Arg	Arg	Asp	Leu 255	Thr
Leu	Gly	Val	Leu 260	Asp	Leu	Val	Ala	Leu 265	Phe	Pro	Ser	Tyr	Asp 270	Thr	Arg
Thr	Tyr	Pro 275	Ile	Asn	Thr	Ser	Ala 280	Gln	Leu	Thr	Arg	Glu 285	Val	Tyr	Thr
Asp	Ala 290	Ile	Gly	Ala	Thr	Gly 295	Val	Asn	Met	Ala	Ser 300	Met	Asn	Trp	Tyr
Asn 305	Asn	Asn	Ala	Pro	Ser 310	Phe	Ser	Ala	Ile	Glu 315	Ala	Ala	Ala	Ile	Arg 320
Ser	Pro	His	Leu	Leu 325	Asp	Phe	Leu	Glu	Gln 330	Leu	Thr	Ile	Phe	Ser 335	Ala
Ser	Ser	Arg	Trp 340	Ser	Asn	Thr	Arg	His 345	Met	Thr	Tyr	Trp	Arg 350	Gly	His
Thr	Ile	Gln 355	Ser	Arg	Pro	Ile	Gly 360	Gly	Gly	Leu	Asn	Thr 365	Ser	Thr	His
Gly	Ala 370	Thr	Asn	Thr	Ser	Ile 375	Asn	Pro	Val	Thr	Leu 380	Arg	Phe	Ala	Ser
Arg 385	Asp	Val	Tyr	Arg	Thr 390	Glu	Ser	Tyr	Ala	Gly 395	Val	Leu	Leu	Trp	Gly 400
Ile	Tyr	Leu	Glu	Pro 405	Ile	His	Gly	Val	Pro 410	Thr	Val	Arg	Phe	Asn 415	Phe
Thr	Asn	Pro	Gln 420	Asn	Ile	Ser	Asp	Arg 425	Gly	Thr	Ala	Asn	Tyr 430	Ser	Gln

Pro	Tyr	Glu 435	Ser	Pro	Gly	Leu	Gln 440	Leu	Lys	Asp	Ser	Glu 445	Thr	Glu	Leu
Pro	Pro 450	Glu	Thr	Thr	Glu	Arg 455	Pro	Asn	Tyr	Glu	Ser 460	Tyr	Ser	His	Arg
Leu 465	Ser	His	Ile	Gly	Ile 470	Ile	Leu	Gln	Ser	Arg 475	Val	Asn	Val	Pro	Val 480
Tyr	Ser	Trp	Thr	His 485	Arg	Ser	Ala	Asp	Arg 490	Thr	Asn	Thr	Ile	Gly 495	Pro
Asn	Arg	Ile	Thr 500	Gln	Ile	Pro	Met	Val 505	Lys	Ala	Ser	Glu	Leu 510	Pro	Gln
Gly	Thr	Thr 515	Val	Val	Arg	Gly	Pro 520	Gly	Phe	Thr	Gly	Gly 525	Asp	Ile	Leu
Arg	Arg 530	Thr	Asn	Thr	Gly	Gly 535	Phe	Gly	Pro	Ile	Arg 540	Val	Thr	Val	Asn
Gly 545	Pro	Leu	Thr	Gln	Arg 550	Tyr	Arg	Ile	Gly	Phe 555	Arg	Tyr	Ala	Ser	Thr 560
Val	Asp	Phe	Asp	Phe 565	Phe	Val	Ser	Arg	Gly 570	Gly	Thr	Thr	Val	Asn 575	Asn
Phe	Arg	Phe	Leu 580	Arg	Thr	Met	Asn	Ser 585	Gly	Asp	Glu	Leu	Lys 590	Tyr	Gly
Asn	Phe	Val 595	Arg	Arg	Ala	Phe	Thr 600	Thr	Pro	Phe	Thr	Phe 605	Thr	Gln	Ile
Gln	Asp 610	Ile	Ile	Arg	Thr	Ser 615	Ile	Gln	Gly	Leu	Ser 620	Gly	Asn	Gly	Glu
Val 625	Tyr	Ile	Asp	Lys	Ile 630	Glu	Ile	Ile	Pro	Val 635	Thr	Ala	Thr	Phe	Glu 640
Ala	Glu	Tyr	Asp	Leu 645	Glu	Arg	Ala	Gln	Glu 650	Ala	Val	Asn	Ala	Leu 655	Phe
Thr	Asn	Thr	Asn 660	Pro	Arg	Arg	Leu	Lys 665	Thr	Asp	Val	Thr	Asp 670	Tyr	His
Ile	Asp	Gln 675	Val	Ser	Asn	Leu	Val 680	Ala	СЛа	Leu	Ser	Asp 685	Glu	Phe	CAa
Leu	Asp 690	Glu	Lys	Arg	Glu	Leu 695	Leu	Glu	Lys	Val	Lys 700	Tyr	Ala	Lys	Arg
Leu 705	Ser	Asp	Glu	Arg	Asn 710	Leu	Leu	Gln	Asp	Pro 715	Asn	Phe	Thr	Ser	Ile 720
Asn	Lys	Gln	Pro	Asp 725	Phe	Ile	Ser	Thr	Asn 730	Glu	Gln	Ser	Asn	Phe 735	Thr
Ser	Ile	His	Glu 740	Gln	Ser	Glu	His	Gly 745	Trp	Trp	Gly	Ser	Glu 750	Asn	Ile
Thr	Ile	Gln 755	Glu	Gly	Asn	Asp	Val 760	Phe	Lys	Glu	Asn	Tyr 765	Val	Thr	Leu
Pro	Gly 770	Thr	Phe	Asn	Glu	Сув 775	Tyr	Pro	Thr	Tyr	Leu 780	Tyr	Gln	Lys	Ile
Gly 785	Glu	Ser	Glu	Leu	Lys 790	Ala	Tyr	Thr	Arg	Tyr 795	Gln	Leu	Arg	Gly	Tyr 800
Ile	Glu	Asp	Ser	Gln 805	Asp	Leu	Glu	Ile	Tyr 810	Leu	Ile	Arg	Tyr	Asn 815	Ala
Lys	His	Glu	Thr 820	Leu	Asp	Val	Pro	Gly 825	Thr	Glu	Ser	Leu	Trp 830	Pro	Leu
Ser	Val	Glu 835	Ser	Pro	Ile	Gly	Arg 840	Сув	Gly	Glu	Pro	Asn 845	Arg	Cys	Ala

Pro	His 850	Phe	Glu	Trp		Pro 855	Asp	Leu	Asp	CÀa	Ser 860	CAa	Arg	l Yab	Gly
Glu 865	Lys	Cys	Ala	His	His 870	Ser	His	His	Phe	Ser 875	Leu	Asp	Ile	e Asp	Val 880
Gly	Сув	Thr	Asp	Leu 885	His	Glu	Asn	Leu	Gly 890	Val	Trp	Val	Val	. Phe	Fys
Ile	Lys	Thr	Gln 900	Glu	Gly	His	Ala	Arg 905	Leu	Gly	Asn	Leu	Glu 910		e Ile
Glu	Glu	Lys 915	Pro	Leu	Leu	Gly	Glu 920		Leu	Ser	Arg	Val 925		arç	, Ala
Glu	Lys 930	Lys	Trp	Arg	-	Lys 935	Arg	Glu	Lys	Leu	Gln 940	Leu	Glu	ı Thı	Lys
Arg 945	Val	Tyr	Thr	Glu	Ala 950	Lys	Glu	Ala	Val	Asp 955	Ala	Leu	Ph∈	· Val	. Asp 960
Ser	Gln	Tyr	Asp	Arg 965	Leu	Gln	Ala	Asp	Thr 970	Asn	Ile	Gly	Met	975	His
Ala	Ala	Asp	980 Lys	Leu	Val	His	Arg	Ile 985	Arg	Glu	Ala	Tyr	Leu 990		Glu
Leu	Pro	Val 995	Ile	Pro	Gly	Val	Asn 100		a Glı	ı Ile	e Ph		u 0	lu I	eu Glu
Gly	His 1010		e Ile	∋ Thi	r Ala	10:		er L	eu T	yr As		la 020	Arg	Asn	Val
Val	Lys 1025		n Gly	Asl	Ph∈	Ası 103		sn G	ly L	eu Tl		ys 035	Trp	Asn	Val
ГÀз	Gly 1040		8 Val	l Asp	o Val	. Gl1		ln S	er H	is H:		rg 050	Ser	Asp	Leu
Val	Ile 1055		Glu	ı Trp	o Glu	106		lu V	al S	er G		la 065	Val	Arg	Val
Cys	Pro 1070		/ Cys	s Gly	y Tyr	10°		eu A:	rg V	al Th		la 080	Tyr	ГÀЗ	Glu
Gly	Tyr 1089	_	/ Glu	ı Gly	у Сув	109		hr I	le H	is G		le 095	Glu	Asn	Asn
Thr	Asp 1100		ı Leı	ı Lys	s Ph∈	Ly:		sn A	rg G	lu G		lu 110	Glu	Val	Tyr
Pro	Thr 1115		Thi	r Gly	y Thr	Cy:		sn A	ap T	yr Tl		la 125	His	Gln	Gly
Thr	Ala 1130		/ Cys	s Ala	a Asp	Ala 113		ys A	sn S	er Ai	-	sn 140	Ala	Gly	Tyr
Glu	Asp 1145		а Туі	r Glu	ı Val	. Ası 119		hr Tl	hr A	la Se		al 155	Asn	Tyr	ГЛа
Pro	Thr 1160		Glu	ı Glu	ı Glu	116		yr Tl	hr A	ab As		rg 170	Arg	Asp	Asn
His	Cys 1179		1 Туі	r Asl	Arg	118		yr V	al A	sn Ty		ro 185	Pro	Val	Pro
Ala	Gly 1190		. Val	l Thi	r Lys	Gl: 119		eu G	lu T	yr Pl		ro 200	Glu	Thr	Aap
Thr	Val 1209	_	) Ile	e Glu	ı Ile	Gly	•	lu Tl	nr G	lu G	•	ys 215	Phe	Ile	Val
Asp	Ser 1220		l Glu	ı Lev	ı Lev	Le:		et G	lu G	lu					

<sup>&</sup>lt;210> SEQ ID NO 50 <211> LENGTH: 1228 <212> TYPE: PRT

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Ser	Leu	Сув 35	Ile	Ala	Glu	Gly	Asn 40	Asn	Ile	Asp	Pro	Phe 45	Val	Ser	Ala
Ser	Thr 50	Val	Gln	Thr	Gly	Ile 55	Asn	Ile	Ala	Gly	Arg 60	Ile	Leu	Gly	Val
Leu 65	Gly	Val	Pro	Phe	Ala 70	Gly	Gln	Leu	Ala	Ser 75	Phe	Tyr	Ser	Phe	Leu 80
Val	Gly	Glu	Leu	Trp 85	Pro	Arg	Gly	Arg	Asp 90	Gln	Trp	Glu	Ile	Phe 95	Leu
Glu	His	Val	Glu 100	Gln	Leu	Ile	Asn	Gln 105	Gln	Ile	Thr	Glu	Asn 110	Ala	Arg
Asn	Thr	Ala 115	Leu	Ala	Arg	Leu	Gln 120	Gly	Leu	Gly	Asp	Ser 125	Phe	Arg	Ala
Tyr	Gln 130	Gln	Ser	Leu	Glu	Asp 135	Trp	Leu	Glu	Asn	Arg 140	Asp	Asp	Ala	Arg
Thr 145	Arg	Ser	Val	Leu	Tyr 150	Thr	Gln	Tyr	Ile	Ala 155	Leu	Glu	Leu	Asp	Phe 160
Leu	Asn	Ala	Met	Pro 165	Leu	Phe	Ala	Ile	Arg 170	Asn	Gln	Glu	Val	Pro 175	Leu
Leu	Met	Val	Tyr 180	Ala	Gln	Ala	Ala	Asn 185	Leu	His	Leu	Leu	Leu 190	Leu	Arg
Asp	Ala	Ser 195	Leu	Phe	Gly	Ser	Glu 200	Phe	Gly	Leu	Thr	Ser 205	Gln	Glu	Ile
Gln	Arg 210	Tyr	Tyr	Glu	Arg	Gln 215	Val	Glu	Arg	Thr	Arg 220	Asp	Tyr	Ser	Asp
Tyr 225	Сла	Val	Glu	Trp	Tyr 230	Asn	Thr	Gly	Leu	Asn 235	Ser	Leu	Arg	Gly	Thr 240
Asn	Ala	Ala	Ser	Trp 245	Val	Arg	Tyr	Asn	Gln 250	Phe	Arg	Arg	Gly	Leu 255	Thr
Leu	Gly	Val	Leu 260	Gly	Leu	Val	Ala	Leu 265	Phe	Pro	Ser	Tyr	Asp 270	Thr	Arg
Thr	Tyr	Pro 275	Ile	Asn	Thr	Ser	Ala 280	Gln	Leu	Thr	Arg	Glu 285	Val	Tyr	Thr
Asp	Ala 290	Ile	Gly	Ala	Thr	Gly 295	Val	Asn	Met	Ala	Ser 300	Met	Asn	Trp	Tyr
Asn 305	Asn	Asn	Ala	Pro	Ser 310	Phe	Ser	Ala	Ile	Glu 315	Ala	Ala	Ala	Ile	Arg 320
Ser	Pro	His	Leu	Leu 325	Asp	Phe	Leu	Glu	Gln 330	Leu	Thr	Ile	Phe	Ser 335	Ala
Ser	Ser	Arg	Trp 340	Ser	Asn	Thr	Arg	His 345	Met	Thr	Tyr	Trp	Arg 350	Gly	His
Thr	Ile	Gln 355	Ser	Arg	Pro	Ile	Gly 360	Gly	Gly	Leu	Asn	Thr 365	Ser	Thr	His
Gly	Ala 370	Thr	Asn	Thr	Ser	Ile 375	Asn	Pro	Val	Thr	Leu 380	Arg	Phe	Ala	Ser
Arg 385	Asp	Val	Tyr	Arg	Thr 390	Glu	Ser	Tyr	Ala	Gly 395	Val	Leu	Leu	Trp	Gly 400

Ile	Tyr	Leu	Glu	Pro 405	Ile	His	Gly	Val	Pro 410	Thr	Val	Arg	Phe	Asn 415	Phe
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Pro	Tyr	Glu 435	Ser	Pro	Gly	Leu	Gln 440	Leu	Lys	Asp	Ser	Glu 445	Thr	Glu	Leu
Pro	Pro 450	Glu	Thr	Thr	Glu	Arg 455	Pro	Asn	Tyr	Glu	Ser 460	Tyr	Ser	His	Arg
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Tyr	Ser	Trp	Thr	His 485	Arg	Ser	Ala	Asp	Arg 490	Thr	Asn	Thr	Ile	Gly 495	Pro
Asn	Arg	Ile	Thr 500	Gln	Ile	Pro	Met	Val 505	ГЛа	Ala	Ser	Glu	Leu 510	Pro	Gln
Gly	Thr	Thr 515	Val	Val	Arg	Gly	Pro 520	Gly	Phe	Thr	Gly	Gly 525	Asp	Ile	Leu
Arg	Arg 530	Thr	Asn	Thr	Gly	Gly 535	Phe	Gly	Pro	Ile	Arg 540	Val	Thr	Val	Asn
Gly 545	Pro	Leu	Thr	Gln	Arg 550	Tyr	Arg	Ile	Gly	Phe 555	Arg	Tyr	Ala	Ser	Thr 560
Val	Asp	Phe	Asp	Phe 565	Phe	Ala	Ser	Arg	Gly 570	Gly	Thr	Thr	Val	Asn 575	Asn
Phe	Arg	Phe	Leu 580	Arg	Thr	Met	Asn	Ser 585	Gly	Asp	Glu	Leu	Lys 590	Tyr	Gly
Asn	Phe	Val 595	Arg	Arg	Ala	Phe	Thr 600	Thr	Pro	Phe	Thr	Phe 605	Thr	Gln	Ile
Gln	Asn 610	Ile	Ile	Arg	Thr	Ser 615	Ile	Gln	Gly	Leu	Ser 620	Gly	Asn	Gly	Glu
Val 625	Tyr	Ile	Asp	Lys	Ile 630	Glu	Ile	Ile	Pro	Val 635	Thr	Ala	Thr	Phe	Glu 640
Ala	Glu	Tyr	Asp	Leu 645	Glu	Arg	Ala	Gln	Glu 650	Ala	Val	Asn	Ala	Leu 655	Phe
Thr	Asn	Thr	Asn 660	Pro	Arg	Arg	Leu	Lys 665	Thr	Asp	Val	Thr	Asp 670	Tyr	His
Ile	Asp	Gln 675	Val	Ser	Asn	Leu	Val 680	Ala	СЛа	Leu	Ser	Asp 685	Glu	Phe	Сув
Leu	Asp 690	Glu	Lys	Arg	Glu	Leu 695	Leu	Glu	Lys	Val	Lys 700	Tyr	Ala	Lys	Arg
Leu 705	Ser	Asp	Glu	Arg	Asn 710	Leu	Leu	Gln	Asp	Pro 715	Asn	Phe	Thr	Ser	Ile 720
Asn	Lys	Gln	Pro	Asp 725	Phe	Ile	Ser	Thr	Asn 730	Glu	Gln	Ser	Asn	Phe 735	Thr
Ser	Ile	His	Glu 740	Gln	Ser	Glu	His	Gly 745	Trp	Trp	Gly	Ser	Glu 750	Asn	Ile
Thr	Ile	Gln 755	Glu	Gly	Asn	Asp	Val 760	Ser	Lys	Glu	Asn	Tyr 765	Val	Thr	Leu
Pro	Gly 770	Thr	Phe	Asn	Glu	Сув 775	Tyr	Pro	Thr	Tyr	Leu 780	Tyr	Gln	Lys	Ile
Gly 785	Glu	Ser	Glu	Leu	Lys 790	Ala	Tyr	Thr	Arg	Tyr 795	Gln	Leu	Glu	Gly	Tyr 800
Ile	Glu	Asp	Ser	Gln 805	Asp	Leu	Glu	Ile	Tyr 810	Leu	Ile	Arg	Tyr	Asn 815	Ala
Lys	His	Glu	Thr	Leu	Asp	Val	Pro	Gly	Thr	Glu	Ser	Leu	Trp	Pro	Leu

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			820					825					830		
Ser	Val	Glu 835	Ser	Pro	Ile	Gly	Arg 840		Gly	Glu	Pro	Asn 845	Arg	Cys	Ala
Pro	His 850	Phe	Glu	Trp	Asn	Pro 855	Asp	Leu	Asp	Cys	Ser 860	CAa	Arg	Asp	Gly
Glu 865	Lys	Сув	Ala	His	His 870	Ser	His	His	Phe	Ser 875	Leu	Asp	Ile	Asp	Val 880
Gly	Сув	Thr	Asp	Leu 885	His	Glu	Asn	Leu	Gly 890	Val	Trp	Val	Val	Phe	
Ile	Lys	Thr	Gln 900	Glu	Gly	His	Ala	Arg 905	Leu	Gly	Asn	Leu	Glu 910	Phe	Ile
Glu	Glu	Lys 915	Pro	Leu	Leu	Gly	Glu 920	Ala	Leu	Ser	Arg	Val 925		Arg	Ala
Glu	Ь 930	Lys	Trp	Arg	Asp	Lys 935	Arg	Glu	Lys	Leu	Gln 940	Leu	Glu	Thr	Lys
Arg 945	Val	Tyr	Thr	Glu	Ala 950	Lys	Glu	Ala	Val	Asp 955	Ala	Leu	Phe	Val	Asp 960
Ser	Gln	Tyr	Asp	Arg 965	Leu	Gln	Ala	Asp	Thr 970	Asn	Ile	Gly	Met	Ile 975	
Ala	Ala	Asp	980 TÀ2	Leu	Val	His	Arg	Ile 985	Arg	Glu	Ala	Tyr	Leu 990	Ser	Glu
Leu	Pro	Val 995	Ile	Pro	Gly	Val	Asn 100		a Glu	ı Ile	e Ph	e Gl		lu L	eu Glu
Gly	His 1010		e Ile	∋ Thi	r Ala	11e		er Le	eu Ty	yr As		la . 020	Arg 2	Asn	Val
Val	Lys 1025		n Gly	y Asp	Phe	Ası 103		sn G	ly Le	eu Th		ys '	Trp 2	Asn	Val
Lys	Gly 1040		s Val	l Ası	Val	Gl1 104		ln Se	er Hi	is Hi		rg 050	Ser I	Asp	Leu
Val	Ile 1055		o Glu	ı Tr <u>ı</u>	Glu	Ala 106		lu Va	al Se	er Gl		la '	Val 1	Arg	Val
CÀa	Pro 1070		y Cys	g Gly	y Tyr	11e		eu Se	er Va	al Th		la 080	Tyr :	ŗàs	Glu
Gly	Tyr 1085		/ Glu	ı Gly	y Cys	Va:		hr I	le Hi	is Gl		le ( 095	Glu Z	Asn	Asn
Thr	Asp 1100	Glu	ı Let		Phe				_	lu Gl			Glu '	Val	Tyr
Pro	Thr 1115		, Thi	r Gly	7 Thr	Cy:		sn As	sp Ty	yr Th		la : 125	His (	Gln	Gly
Thr	Ala 1130		y Cys	s Alá	a Asp	Ala 113		ys As	sn Se	er Ai		sn . 140	Ala	Gly	Tyr
Glu	Asp 1145		а Туг	r Glu	ı Val	As)		hr Tl	nr Al	la Se		al . 155	Asn '	Tyr	Lys
Pro	Thr 1160	_	r Glu	ı Glı	ı Glu	Th:		yr Tl	nr As	∍p Va		rg . 170	Arg 2	Asp	Asn
His	Cys		а Туг	r Asl	Arg	Gl <sub>3</sub>		yr Va	al As	∍n Ty		ro :	Pro '	Val	Pro
Ala	Gly 1190	_	r Val	l Thi	r Lys	Gl:		eu G	lu Ty	yr Pl		ro (	Glu '	Thr	Asp
Thr	Val 1205		o Ile	e Glu	ı Ile	Gl <sub>3</sub>		lu Tì	nr Gl	lu Gl		ys : 215	Phe l	Met	Val
Asp		Va]	l Gli	ı Lev	ı Leu		a M	et G	lu G	lu					

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Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile I 50 60	le
Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln I 65 70 75 8	
Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln A 85 90 95	la
Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala G 100 105 110	lu
Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg G 115 120 125	lu
Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr A	la
Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser V. 145 150 155 1	al 60
Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val S 165 170 175	er
Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser A	rg
Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp His Ala V 195 200 205	al
Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser A	rg
Asp Trp Ile Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr V. 225 230 235 25	al 40
Leu Asp Ile Val Ser Leu Phe Pro Asn Tyr Asp Ser Arg Thr Tyr Program 245 250 250 255	ro
Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro V. 260 265 270	al
Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile G 275 280 285	lu
Gly Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile T 290 295 300	hr
Ile Tyr Thr Asp Ala His Arg Gly Glu Tyr Tyr Trp Ser Gly His G 305 310 315 3.	ln 20
Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Ph 325 330 335	ro
Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val A 340 345 350	la
Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr A 355 360 365	rg
Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu A	ap

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Tyr	Arg	Lys	Ser	Gly 405	Thr	Val	Asp	Ser	Leu 410	Asp	Glu	Ile	Pro	Pro 415	Gln
Asn	Asn	Asn	Val 420	Pro	Pro	Arg	Gln	Gly 425	Phe	Ser	His	Arg	Leu 430	Ser	His
Val	Ser	Met 435	Phe	Arg	Ser	Gly	Phe 440	Ser	Asn	Ser	Ser	Val 445	Ser	Ile	Ile
Arg	Ala 450	Pro	Met	Phe	Ser	Trp 455	Ile	His	Arg	Ser	Ala 460	Glu	Phe	Asn	Asn
Ile 465	Ile	Pro	Ser	Ser	Gln 470	Ile	Thr	Gln	Ile	Pro 475	Leu	Thr	ГÀа	Ser	Thr 480
Asn	Leu	Gly	Ser	Gly 485	Thr	Ser	Val	Val	Lys 490	Gly	Pro	Gly	Phe	Thr 495	Gly
Gly	Asp	Ile	Leu 500	Arg	Arg	Thr	Ser	Pro 505	Gly	Gln	Ile	Ser	Thr 510	Leu	Arg
Val	Asn	Ile 515	Thr	Ala	Pro	Leu	Ser 520	Gln	Arg	Tyr	Arg	Val 525	Arg	Ile	Arg
Tyr	Ala 530	Ser	Thr	Thr	Asn	Leu 535	Gln	Phe	His	Thr	Ser 540	Ile	Asp	Gly	Arg
Pro 545	Ile	Asn	Gln	Gly	Asn 550	Phe	Ser	Ala	Thr	Met 555	Ser	Ser	Gly	Ser	Asn 560
Leu	Gln	Ser	Gly	Ser 565	Phe	Arg	Thr	Val	Gly 570	Phe	Thr	Thr	Pro	Phe 575	Asn
Phe	Ser	Asn	Gly 580	Ser	Ser	Val	Phe	Thr 585	Leu	Ser	Ala	His	Val 590	Phe	Asn
Ser	Gly	Asn 595	Glu	Val	Tyr	Ile	Asp 600	Arg	Ile	Glu	Phe	Val 605	Pro	Ala	Glu
Val	Thr 610	Phe	Glu	Ala	Glu	Tyr 615	Asp	Leu	Glu	Arg	Ala 620	Gln	ГÀЗ	Ala	Val
Asn 625	Glu	Leu	Phe	Thr	Ser 630	Ser	Asn	Gln	Ile	Gly 635	Leu	Lys	Thr	Asp	Val 640
Thr	Asp	Tyr	His	Ile 645	Asp	Gln	Val	Ser	Asn 650	Leu	Val	Glu	CÀa	Leu 655	Ser
Asp	Glu	Phe	660 CÀa	Leu	Asp	Glu	Lys	Lys 665	Glu	Leu	Ser	Glu	Lys 670	Val	Lys
His	Ala	Lys 675	Arg	Leu	Ser	Asp	Glu 680	Arg	Asn	Leu	Leu	Gln 685	Asp	Pro	Asn
Phe	Arg 690	Gly	Ile	Asn	Arg	Gln 695	Leu	Asp	Arg	Gly	Trp 700	Arg	Gly	Ser	Thr
Asp 705	Ile	Thr	Ile	Gln	Gly 710	Gly	Asp	Asp	Val	Phe 715	Lys	Glu	Asn	Tyr	Val 720
Thr	Leu	Leu	Gly	Thr 725	Phe	Asp	Glu	Cys	Tyr 730	Pro	Thr	Tyr	Leu	Tyr 735	Gln
Lys	Ile	Asp	Glu 740	Ser	Lys	Leu	Lys	Ala 745	Tyr	Thr	Arg	Tyr	Gln 750	Leu	Arg
Gly	Tyr	Ile 755	Glu	Asp	Ser	Gln	Asp 760	Leu	Glu	Ile	Tyr	Leu 765	Ile	Arg	Tyr
Asn	Ala 770	Lys	His	Glu	Thr	Val 775	Asn	Val	Pro	Gly	Thr 780	Gly	Ser	Leu	Trp
Pro 785	Leu	Ser	Ala	Pro	Ser 790	Pro	Ile	Gly	Lys	Суs 795	Ala	His	His	Ser	His 800

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Н	is	Phe	Ser	Leu	Asp 805	Ile	Asp	Val	Gly	Cys 810	Thr	Asp	Leu	Asn	Glu 815	_
L	eu	Gly	Val	Trp 820	Val	Ile	Phe	Lys	Ile 825		Thr	Gln	Asp	Gly 830		Ala
A	rg	Leu	Gly 835	Asn	Leu	Glu	Phe	Leu 840	Glu	Glu	Lys	Pro	Leu 845		Gly	Glu
A	la.	Leu 850	Ala	Arg	Val	Lys	Arg 855	Ala	Glu	Lys	Lys	Trp 860	Arg	Asp	Lys	Arg
	lu 65	Lys	Leu	Glu	Trp	Glu 870	Thr	Asn	Ile	Val	Tyr 875	-	Glu	Ala	Lys	Glu 880
S	er	Val	Asp	Ala	Leu 885	Phe	Val	Asn	Ser	Gln 890	Tyr	Asp	Arg	Leu	Gln 895	
A	.sp	Thr	Asn	Ile 900	Ala	Met	Ile	His	Ala 905	Ala	Asp	ГÀз	Arg	Val 910		Ser
Ι	le	Arg	Glu 915	Ala	Tyr	Leu	Pro	Glu 920	Leu	Ser	Val	Ile	Pro 925	_	Val	Asn
A	la.	Ala 930	Ile	Phe	Glu	Glu	Leu 935	Glu	Gly	Arg	Ile	Phe 940	Thr	Ala	Phe	Ser
	eu 45	Tyr	Asp	Ala	Arg	Asn 950	Val	Ile	Lys	Asn	Gly 955	Asp	Phe	Asn	Asn	Gly 960
			-		965			Gly		970					975	
				980				Val	985					990		
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		1085	5				109					1	095			
		1100	)				110					1	110			
		1115	5				112					1	125			
		1130	)				113			-		1	140	•		
		1145	-				11!						155			

The invention claimed is:

- 1. An engineered Cry1Ba (eCry1Ba) protein comprising a mutation at a position corresponding to amino acids 2 and 150; or amino acids 2, 150 and 178; or amino acids 2, 150 and 189; or amino acids 2, 150 and 199, of SEQ ID NO: 5, said protein having improved solubility or insecticidal activity
- against at least European corn borer (*Ostrinia nubilalis*) when compared to a native or wild-type Cry1Ba protein.
- 2. The eCry1Ba protein of claim 1, wherein the mutation is at amino acids 2 and 150; or amino acids 2, 150 and 178; or amino acids 2, 150 and 189; or amino acids 2, 150 and 199, of SEQ ID NO: 5.

- 3. The eCry1Ba protein of claim 1, wherein the amino acid corresponding to position 2 is any amino acid and
  - (a) the amino acid corresponding to position 150 is Lys; or
  - (b) the amino acid corresponding to position 150 is Lys and the amino acid corresponding to position 178 is Ser; or
  - (c) the amino acid corresponding to position 150 is Lys and the amino acid corresponding to position 189 is Ser; or
  - (d) the amino acid corresponding to position 150 is Lys and the amino acid corresponding to position 199 is Lys.
- **4**. The eCry1Ba protein of claim **3**, wherein the protein comprises an amino acid sequence of SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9 or SEQ ID NO: 10.
- **5.** The eCry1Ba protein of claim **1**, wherein the protein has insecticidal activity against an additional lepidopteran insect selected from the group consisting of southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean caterpillar.
- **6**. The eCry1Ba protein of claim **1**, wherein the specific activity of said eCry1Ba protein is at least  $3 \times$  higher than a native Cry1Ba protein.
- 7. A Cry1Ba protein comprising SEQ ID NO: 3 or SEQ ID NO:11 or SEQ ID NO:12.
- **8**. The Cry1Ba protein of claim **7**, wherein the protein has activity against a lepidopteran insect.
- **9.** The Cry1Ba protein of claim **8**, wherein the lepidopteran <sup>25</sup> insect is European corn borer, southwestern corn borer or sugarcane borer.
- 10. An insecticidal composition comprising the engineered Cry1Ba protein of claim 1 and an acceptable agricultural carrier.
- 11. The composition of claim 10, wherein the agricultural carrier is a plant which expresses the eCry1Ba protein.
- 12. A method of making an eCry1Ba protein comprising: (a) identifying a Cry1Ba protein having a domain I of said protein; (b) mutating the Cry1Ba protein at a position corresponding to amino acids 2 and 150; or amino acids 2, 150 and 178; or amino acids 2, 150 and 189; or amino acids 2, 150 and 199, of SEQ ID NO: 5; and (c) obtaining the eCry1Ba protein

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so produced, wherein the eCry1Ba has improved solubility or insecticidal activity against at least European corn borer when compared to a native Cry1Ba protein.

- 13. The method of claim 12, wherein the mutation is at amino acids 2 and 150; or amino acids 2, 150 and 178; or amino acids 2, 150 and 189; or amino acids 2, 150 and 199, of SEO ID NO: 5.
- 14. The method of claim 12, wherein the mutation at position 2 is any amino acid and
  - (a) the amino acid corresponding to position 150 is Lys; or (b) the amino acid corresponding to position 150 is Lys and the amino acid corresponding to position 178 is Ser; or
  - (c) amino acid corresponding to position 150 is Lys and the amino acid corresponding to position 189 is Ser; or
  - (d) the amino acid corresponding to position 150 is Lys and the amino acid corresponding to position 199 is Lys.
- **15**. The method of claim **14**, wherein the eCry1Ba protein comprises an amino acid sequence of SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9 or SEQ ID NO: 10.
- 16. The method of claim 12, wherein the eCry1Ba protein has activity against lepidopteran or coleopteran insects.
- 17. The method of claim 16, wherein the lepidopteran insects are European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean caterpillar.
- 18. A method of controlling a lepidopteran insect comprising contacting the insect with an effective amount of an eCry1Ba protein having a mutation at a position corresponding to amino acids 2 and 150; or amino acids 2, 150 and 178; or amino acids 2, 150 and 189; or amino acids 2, 150 and 199, of SEQ ID NO: 5, the protein having improved solubility or insecticidal activity against a lepidopteran insect when compared to a native Cry1Ba protein.
- 19. The method of claim 18, wherein the lepidopteran insect is selected from the group consisting of a European corn borer, southwestern corn borer, sugarcane borer, corn earworm, soybean looper and velvet bean caterpillar.

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